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The influence of the composition of the nutrient medium on the morphogenesis of *Rhododendron luteum* Sweet, introduced varieties of *Vaccinium corymbosum* L. and *Vaccinium vitis-idaea* L.

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Abstract

The morphogenesis of the introduced varieties of *Vaccinium corymbosum*, *Vaccinium vitis-idaea*, *Rhododendron luteum* on various modifications of the nutrient media was studied, the optimal composition of the nutrient medium for this process was determined. The principal possibility of regeneration of introduced varieties of *Vaccinium corymbosum*, *Vaccinium vitis-idaea* by the method of activation of axillary meristems was shown; for *Rhododendron luteum*— by two methods: 1) by activation of the axillary meristems, 2) through the proliferation of callus and the subsequent formation of shoots from it.

Keywords: morphogenesis, nutrient media, introduced varieties

Introduction

Extensive literature is devoted to the question of morphogenesis in the culture of cells and tissues. Its analysis allows us to conclude that morphogenesis is a complex and multifactorial process, depending on the type and physiological state of the explant, composition of the nutrient medium, i.e. the components, contained in it (macromicroelements, vitamins, carbohydrates, hormonal supplements), as well as on the pH of the medium. cultivation conditions and a number of other factors. This is confirmed by numerous experimental studies.

Gupta and Chandra (1985) studied the effect of various growth regulators (BAP, NAA, GA) on the morphogenesis of various types of tobacco explants:

leaf pieces without a central vein, isolated from 2–4 upper leaves; segments of internodes isolated from the second upper internodes; stripes of epidermal tissue with several adjacent layers of cells isolated from young internodes. Experimental data allowed the authors to conclude that GA in a concentration of 0.5 mg / L stimulated the formation of buds only on explants of leaf pieces; kinetin and NAA promoted to the formation of vegetative buds on stem explants, and kinetin on leaf explants.

According to the results of studies by Shor and Papazyan(1989), obtained by studying the processes of morphogenesis in a culture of isolated rose tissues on five media, differing in the concentration of macrosalts and a combination of hormonal supplements, the realization of morphogenesis

consisted in the development of shoots from axillary buds and the formation of callus on sections of the stem and petiole of the leaf. The most intensive shoot development was noted on the Murashige-Skoog medium of full mineral composition with the addition of BAP and NAA under a 16-hour photoperiod.

From a publication by Vilor et al. (1987), it follows, that the morphogenetic processes, occurring in sunflower in an in vitro culture depend on the type of nutrient medium and explant. They found, that callus was formed best of all on Erickson and Murashige-Skoog media from the apical meristem of stem, and on the medium of White - from a leaf. The authors observed the formation of shoots with roots only from the apical meristem.

The role of auxins and cytokinins in the regulation of morphogenesis is evidenced by experimental studies by Budagovskava et al. (1990). The leaves and tops of young shoots of cereals grown under aseptic conditions, as well as the leaves of adult plants cultivated in the field, were used as explants. The authors conclude that calliare formed better on explants, taken from adult plants, grown in the field, with 1 mg / L benzyladenine and 1.2 NAA in the medium. Shoot formation was noted on Murashige-Skoog medium, containing 2 mg / 1 benzyladenine.

A study of the morphogenesis of introduced varieties of *Vaccinium corymbosum*, *Vaccinium vitis-idaea*, *Rhododendron luteum*, on various modifications of the nutrient media, will determine the optimal composition of the nutrient medium for passing of this physiological process in vitro.

Materials and Methods

The objects of study were introduced varieties of *Vaccinium corymbosum* 'Elizabeth', *Vaccinium vitisidaea*' Ammerland', 'Red Pearl', *Rhododendron luteum*. The experiments were performed on three types of nutrient media:MS (Murashige and Skoog, 1962), WPM (Lloyd and McCown, 1981), Anders (Anderson, 1975), represented by 9 different modifications (Table 1).

As explants there were us d micrografts of introduced varieties of *Vaccinium corymbosum* 'Elizabeth', *Vaccinium vitis-idaea*' Ammerland', 'Red Pearl', *Rhododendron luteum*, introduced into a sterile culture, and also epicotyl, hypocotyl, cotyledon, root, leaves of juvenile seedlings of *Rhododendron luteum*,

obtained by us earlier under aseptic conditions on modified Anders n s nutrient medium. Sterile explants were planted on nutrient media: Murashige-Sk og, WPM and Anders n in flasks of the same volume of 15 ml of medium in each. The planted material was cultivated at a temperature of 26 ° C, humidity of 56%, a photoperiod of 16 hours, and illumination of 4,000 lux. The repetition of experiments was three times. The number of shoots per explant (pcs.), callus formation (mg) after 45 days from the moment of planting of explants on a nutrient medium was taken into account. Statistical data processing was carried out on the basis of 20 explants for repetition. The experimental data are summarized in the table. 2-3. They show arithmetical means and their standard errors.

Results and Discussion

After four weeks of cultivation from one micrograft was formed on the average of 1 to 13 microshoots, depending on the composition of the nutrient medium (table 2). In explants of Rhododendron luteum (epicotyl, hypocotyl, cotyledons, root, leaves), after 5-6 weeks of cultivation, organogenic callus was formed with subsequent regeneration of vegetative shoots from it. It should be noted that the formation of organogenic callus and the subsequent regeneration of shoots are typical for explants (root, epicotyl, hypocotyl, cotyledons, leaves,) obtained from freshly harvested seeds, and for explants from germinated seeds that have passed stratification, shoot formation occurred directly from explant tissue, by passing the stage of callus formation. It is logical to assume, that this may be due to the unequal course of physiological, biochemical, cytological and other processes in explants from freshly harvested and stratificated seeds, as well as with a different content of endogenous phytohormones in them. Probably, all taken together served as the basis for the regeneration of shoots from callus without preliminary passaging it on a nutrient medium of other composition. In other words, the induction of callusogenesis, and then the formation of shoots, took place on a medium of the same composition.

From table 3 it follows that the highest morphogenetic potential is possessed by all explants of *Rhododendron luteum* on media: WPM and Anders n of two modifications (No. 8, 9, see table 1). In this case, the morphogenesis of *Rhododendron luteum* is based on the ability of explant cells to be dedifferentiated, in other words, to lose their previous specialization and turn into callus cells. The transformation of

Table 1- Composition of nutrient media for studying the morphogenesis of introduced varieties of Vaccinium corymbosum, Vaccinium vitis-idaea, Rhododendron luteum

Component, mg/l	Modification of medium								
	1	2	3	4	5	6	7	8	9
Salts and vitamins on S	+	-	1/2	+	-	-	-	-	-
Salts and vitamins on WPM	-	+	-	-	-	-	-	+	-
Salts and vitamins on Andersen	-	-	-	-	+	+	+	-	+
Mesoinositol	100	100	100	100	100	100	100	100	100
Adenine sulphate	-	80	80	80	80	40	60	80	80
Thiamine	0,4	-	-	0,4	-	0,1	0,1	0,4	0,1
Pyridoxine	-	-	-	0,4	-	-	-	-	-
Indolylacetic acid	1,0	5,0	-	2,0	2,0	1,5	2,5	4,0	4,0
Gibberelic acid	-	4,0	-	-	-	-	-	-	-
Naphtylacetic acid	-	-	-	-	-	-	-	-	-
Benzylaminopurine	-	-	-	-	-	2,0	-	-	-
Isopenteniladenine	10	10	2,0	5,0	4,0	-	10	15	15
Saccharose, g/l	20	20	20	30	30	20	20	30	30
Agar, g/l	9	9	9	9	9	9	9	9	9
	4,8	4,8	4,8	4,8	4,0	4,0	4,0	4,8	4,8

Notation: (+) – component is present in the medium; (-) –component is absent in the medium; ½ –half dose of the component in the medium.

Table 2- Shoot formation in introduced varieties of *Vaccinium corymbosum*, *Vaccinium vitis-idaea*, *Rhododendron luteum*, depending on the composition of the nutrient medium

	Quantity of regenerants on one explant, piece						
Number of modification of medium	'Elizabeth'	'Ammerland'	'Red Pearl'	Rhododendron luteum			
1	4,5±1,2	$4,7\pm1,8$	4,2±1,0	4,5±1,3			
2	$3,5\pm1,4$	$3,2\pm1,0$	$3,7\pm1,2$	$4,2\pm1,0$			
3	$1,1\pm1,0$	$1,3\pm1,0$	$1,8\pm0,2$	1,3±1,0			
4	1,2±1,3	$2,9\pm1,1$	$3,2\pm1,1$	$3,1\pm1,7$			
5	$3,6\pm1,2$	$3,1\pm1,3$	$3,0\pm 2,1$	$2,1\pm1,2$			
6	1,3±1,1	$0,7\pm0,1$	1,0±0,3	$0,6\pm0,1$			
7	$1,4\pm1,0$	$1,6\pm1,1$	$1,2\pm0,1$	1,5±1,0			
8	7,0±1,0	$10,0\pm1,0$	$11,5\pm2,1$	6,0±1,7			
9	9,0±1,0	12,0±2,0	13,0±2,0	7,0±2,2			

Table 3 – Morphogenesis in *Rhododendron luteum*, depending on the composition of the nutrient medium.

	Quantity of regenerants on one explant, piece								
Number of	callus, mg	shoots, piece.	Source of explants						
modification of medium			rootlet	hypocotyl	epicotyl	cotyledons	leaves		
1	30,7±3,1	1,0±0,0	+	+	+	+	+		
2	165,6±3,8	$10,0\pm3,0$	++	++	++	++	++		
3	130,0±3,2	$9,0\pm1,0$	++	++	++	++	++		
4	210,0±3,0	$16,0\pm1,0$	+++	+++	+++	+++	+++		
5	110,5±16,1	$13,0\pm2,0$	+++	+++	+++	+++	++++		
6	$40,8\pm1,4$	$2,0\pm1,0$	+	+	+	+	+		
7	85,0±2,5	$7,0\pm 2,0$	+	+	+	+	+		
8	119,0±1,7	$8,0\pm2,0$	++	++	++	++	++		
9	305,0±6,1	19,0±3,0	+++	+++	+++	+++	+++		

Notation: (+) – morphogenesis is low, (++) –average morphogenesis, (+++) –high morphogenesis

specialized cells into callus cells is associated with the induction of cell division, the ability of which cells lost in the process of differentiation (Butenko,1975).

According to the theory of Skoog and Miller (1957), the process of morphogenesis begins from the transition of the cell to the initiation of organized development and is the result of a change in the balance between phytohormones. They found, that the excess of auxin content over cytokinin in the medium causes the induction of roots; inverse correlation i.e. the excess of cytokinin over auxin leads to the formation of buds and stem shoots.

It can be assumed that differences between cells and tissues in the content of endogenous phytohormones determine the different character of their behavior in an isolated culture and the unequal needs in components of medium.

Callus cells (with the exception of auxin- and cytokinin-independent tumor cells) cannot synthesize phytohormones themselves in sufficient quantities necessary for the induction of morphogenesis, therefore, they need exogenous growth regulators. Callus cells only with a certain ratio of cytokinins and auxins in the medium can go on to organized growth and shoot formation. This ratio for every species of plant is established experimentally. This can be confirmed by numerous studies on the regulation of morphogenesis in cell and tissue culture using a certain ratio of auxins and cytokinins in a nutrient medium (Grozeva and Nankar, 2020; Sharaf et al., 2011; Mohamed and Alsadon, 2011; Abbas and Qaiser, 2012; Dadvar et al., 2013; Kakarla et al., 2014; Ngobile et al., 2015; Christopher et al.,1987; Makoveychuk, 1990; Cheruvathur et al., 2015; Hala and Almobasher, 2016; Sweety and Rahman, 2016; Ali et al., 2017; Shete et al., 2017; Kaveri and Srinath, 2017; Yandia et al., 2018; Fikadu and Tileye, 2019; Khan et al., 2019; Choudhari et al., 2020; Dereje et al., 2020).

Our studies have shown, that for the formation regenerants of *Rhododendron luteum* from callus tissue into nutrient medium, it is necessary to add cytokinins and auxins in the following ratios: 2.5: 1 (m dium No. 4), 2: 1 (medium No. 5), 3.75: 1 (medium No. 8 and No. 9).

As shown by the analysis of the results of experimental studies on the morphogenesis of introduced varieties of *Vaccinium corymbosum*, *Vaccinium vitis-idaea*, *Rhododendron luteum* on nine

modifications of nutrient media, that differ in the content of macro- and microsalts, hormonal supplements, best for morphogenesis of studied plants were the 8th and 9th modifications, containing macro- and microelements of WPM and Anderson media, as well as hormonal supplements: 4 mg / 1 indolylacetic acid and 15 mg / 1 isopentenyladenine (Table 1). On media of the 8th and 9th modifications, in comparison with those of the 1st, 2nd, 3rd, 4th, 5th, 6th and 7th, the maximum number of shoots per explant was obtained from 6 to 13 depending on the variety and type of plant (Table 2).

Conclusion

The best for morphogenesis of introduced varieties of *Vaccinium corymbosum*, *Vaccinium vitis-idaea* and *Rhododendron luteum* were media of the 8th and 9th modifications, containing macro- and microelements of WPM and Andersen media, as well as hormonal supplements: 4 mg / 1 indolylacetic acid and 15 mg / 1 isopentenyladenine. The principle possibility of regeneration of *Rhododendron luteum* by two methods has been shown: 1) by activating the axillary meristems, 2) through the proliferation of callus and the subsequent formation of regenerants from it.

References

- Abbas, H., Qaiser, M. 2012. In vitro response of *Ruellia bracteolata* to different growth hormones—an attempt to conserve an endangered species. Plant Cell Tiss. Organ Culture. 44(2): 791-794.
- Ali, J., Bantte, K., Feyissa, T. 2017.Protocol optimization for *in vitro* shoot multiplication of Jackfruit (*Artocarpus heterophyllus* L.).African Journal of Biotechnology.16 (2): 87-90.
- nderson, W.C. 1975.Propagation of rhododendrons by tissue culture. Part1. Development of culture medium for multiplication of shoots. Proc. Intern. Plant Prop. Soc. 25:1929-1935.
- Budagovskaya, N.V., Kara, A.N., Kotov, A.A.1990. Hormonal regulation of pea, isolated apex development. Plant Physiol. 79 (2), pt. 2: 7.
- Butenko, R.G. 1975. Experimental morphogenesis and differentiation in culture of cells of plants. Moscow: Nauka: 51.
- Cheruvathur, M. K., Abraham, J., Thomas, T. D. 2015. *In vitro* micropropagation and flowering in *Ipomoea sepiaria* Roxb. An important ethanomedicinal plant Asian Pacific. Journal of Reproduction.4(1): 49-53.

- Choudhari, N.B., Khade, R.S., Thakare, I.S. 2020. In vitro Medicinal plant *Uraria picta* Jacq DC. Int. J. Adv. Res. Biol. Sci. 7 (4): 169-172.
- Christopher, T., Prolaram, B., Rajam, M., Subhash, V. 1987. In vitro response of excised embryos from red pepper (*Capsicum annuum* L.) on hydroxylamine treatment. Indian. J. Exp. Biol. 25(5): 349-350.
- Dadvar, F., Shahraji, T.R., Assare, M.H., Emam, M., Shirvany, A. 2013. Effects of different concentrations of plant regulators on in vitro micropropagation of *Celtis caucasica* Willd. Iranian Journal of Rangelands and Forests Plant Breeding and Genetic Research. 21(1): 13-22.
- Dereje, H., Buko, T.,Hvoslef-Eide,A. K. 2020. Optimization of plant growth regulators for meristem initiation and subsequent multiplication of five virus tested elite sweet potato varieties from Ethiopia. African Journal of Biotechnology. 19(6): 332-343.
- Fikadu, K., Tileye, F. 2019. In vitro regeneration of two grapevine (*Vitis vinifera* L.) varieties from leaf explants. African Journal of Biotechnology. 18(4): 92-100.
- Grozeva, S., Nankar, A.N. 2020. Effect of incubation period and culture medium on pepper anther culture. Indian Journal of Biotechnology. 19(1):53-59.
- Gupta,S.C., Chandra, N.1985. Control of organogenesis in cultures of different vegetative explants of *Nicotian plumbaginifolia* Viv.Indian.J. Plant. Physiol. 2: 145-150.
- Hala, Al., Almobasher, A. 2016. Comparison Study On *In Vitro* morphogenesis of Mature and Immature Wheat (*Triticum aestivum* L.) Embryos. International Journal of Advanced Biotechnology and Research. 7 (3): 1134-1141.
- Kakarla, L., Rama, C., Botlagunta, M., Krishna, M.S., Mathi, P.S. 2014. Somatic embryogenesis and plant regeneration from leaf explants of *Rumex vesicarius* L. African Journal of Biotechnology. 13(45): 4268-4274.
- Kaveri, S., Srinath, R. 2017. Thidiazuron mediated callus and multiple shoot induction in *Nothapodytes foetida* (Wight) Sleumer an important medicinal plant. International Journal of Current Advanced Research. 6 (2):1731-1734.
- Khan, T. A., Pathak, D., Sharma, P., Ansari, M., Agnihotri, R. K. 2019. In vitro regeneration, callus induction and rhizogenesis in *Ficus krishnae*: A rare endangered plant. Indian Journal of Biotechnology. 18(4):346-351.

- Lloyd, G., McCown. 1981. Commercially feasible micropropagation of mountain laurel, *Kalmia latifolia*, by use of shoot tip culture. Proc. Intern. Plant Prop. Soc. 30:421-427.
- Makoveychuk, A.Y.1990. Embryogenesis as a model of correlative interaction of phytohormones. The Second All-Union Congress of the Society of Plant Physiologists: Proceedings of the International Scientific Conference, Minsk, September 24-29:58.
- Mohamed, M.A., Alsadon, A.A.2011. Effect of vessel type and growth regulators on micropropagation of *Capsicum annuum*. Biologia Plantarum. 55 (2): 370–374.
- Murashige, T., Skoog, F. 1962.A revised medium for rapid growth and bioassays with tobacco tissue cultures. Physiol. Plant. 15: 473-497.
- Nqobile,M.,Adeyemi,A.,Jeffrey,F.,Johannes,S.2015.G rowth and phytochemical levels in micropropagated *Eucomis autumnalis* (Mill.)Chitt. Subspecies autumnalis using different gelling agents, explant source, and plant growth regulators. *In Vitro* Cellular & Developmental Biology Plant. 51(1):102-110.
- Sharaf, A.R., Hamidoghli, Y., Zakizadeh, H.2011. In vitro Seed Germination and Micropropagation of Primrose (*Primula heterochroma* Stapf.) an Endemic Endangered Iranian Species via Shoot Tip Explants.Horticulture, Environment and Biotechnology.52 (3): 298–302.
- Shete, R., Jadhav, A., Pandhure, N. 2017. *In vitro* multiplication of *Solanum virginianum* L. Int. J. Adv. Res. Biol. Sci. 4 (2): 157-160.
- Shor, M.F., Papazian, N.D. 1989.Study of the processes of morphogenesis in culture of isolated tissues roses. Rus. acad. of sciences. Inst of Plant Physiology, Moscow, Dep. VINITI 19.04.89. No 2572-889.
- Skoog, F., Miller, C.O. 1957. Chemical regulation of growth and organ formation in plant tissues cultured in vitro. In: The biological action of growth substances: Symp. Soc. Exp. Biol. Cambridge.11: 118-123.
- Sweety, M.,Rahman, M. 2016. In Vitro Rapid Clonal Propagation of *Plumbago zeylanica* Linn. Through Direct Organogenesis. International Journal of Advanced Biotechnology and Research.7(3): 877-887.
- Vilor, T.A., Gaponenko, A.K., Melkonov, N.M.1987. Selection of the optimal nutrient medium for sunflower. Rus.acad.of sciences. Inst of Plant Physiology. M., Dep. VINITI 19.01.87. No 328-387.

Yandia, S.P., Gandonou, C.B.,Silla, S., Zinga, I., Toukourou, F. 2018.Response of four cultivars of cassava (*Manihot esculenta* Crantz) plantlets free of cassava mosaic virus to micropropagation in different media. African Journal of Biotechnology. 17(1): 9-16.



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