



Green synthesis, characterization and antibacterial activity of iron oxide nanoparticles derived from *Solanum nigrum* leaf extract

M. Rashmi Sulthana¹ and Dr. M. Shenbagam Madhavan^{2*}

¹Faculty of Science, Department of Biochemistry and Biotechnology, Annamalai University, Annamalainagar, Tamilnadu- 608002, India.

^{2*}Assistant Professor, Faculty of Science, Department of Biochemistry and Biotechnology, Annamalai University, Annamalainagar, Tamilnadu- 608002, India.

Corresponding Author E-mail: shenbu15@gmail.com

Abstract

In order to create iron nanoparticles, extract from *Solanum nigrum* was used. By using UV/Vis absorption spectroscopy, X-ray diffraction spectroscopy, and SEM examination, the produced iron nanoparticles were identified. Iron oxide nanoparticles exhibit a distinctive absorption peak in the UV/Vis spectrum between 200 and 300 nm. The average particle size of magnetite nanoparticles, as determined by the X-ray diffraction method, was found to be 24.1 nm. By using the well diffusion method, it was discovered that the synthetic iron nanoparticles have antibacterial action against harmful bacteria such *Staphylococcus aureus*, *Streptococcus pyogenes*, *Escherichia coli*, and *Klebsiella pneumoniae*. This biosynthetic method has been discovered to be economical, environmentally benign, and promising for use in a variety of fields.

Keywords: Iron nanoparticles, *Solanum nigrum*, antibacterial activity

Introduction

Utilizing materials at the nanoscale, or with a dimension smaller than 100 nm, is the subject of nanotechnology. The produced nanoparticles have particular features and are used in a variety of applications, including biological labels, sensors, health care devices, data storage, catalysis, pigment, and ion exchangers [1,2]. These nanoparticles are used in a variety of fields, such as food and agriculture, environment, medicine, and so on[3-5]. They have special physical,

optical, chemical, electrical, and medical properties. Due to their superparamagnetic properties and employment as contrast agents in nuclear magnetic resonance, magnetic nanoparticles have recently become a major concern in the healthcare sector. They are also utilised for cancer hyperthermia treatment, sustained and targeted medication administration, etc. Utilizing materials at the nanoscale, or with a dimension smaller than 100 nm, is the subject of nanotechnology. They are also employed in cancer hyperthermia therapy, sustained and

targeted drug delivery, and other applications. The two main requirements for a substance to be used in medicine are that it must be non-toxic and biocompatible. This section is very concerned with iron oxide nanoparticles, which are an inorganic transition metal oxide. Iron oxide nanoparticles have been created using a variety of techniques. This covers both physical and chemical procedures such as Co-Precipitation, Thermal Decomposition, Microemulsion, Hydrothermal Synthesis, Sonochemical Synthesis, Microwave Method, etc. [6,8]

However, these techniques have a number of drawbacks, including high costs, slow production rates, hazardous chemicals, and by product contamination. A straightforward, effective, and environmentally friendly solution is needed to address this. This is made possible by the new "Green Chemistry" approach, which uses a safe and non-toxic process to create nanoparticles. Microbes and plant extracts are employed for this [9–11]. Plant extracts are chosen over microorganisms for the creation of nanoparticles because they may be produced more quickly and require less upkeep of cell cultures [12]. In the current study, iron oxide is synthesized in a green manner using a leaf extract from *Solanum nigrum* plants.

Materials and Methods

Synthesis of Fe₂O₃ nanoparticle

Preparation of *Solanum nigrum* leaf extract

Solanum nigrum leaves weighing about 25g were obtained from the market in Chidambaram, Tamil Nadu. They were properly cleaned with distilled water, chopped into little pieces, and then cooked in 100 mL of the same water until the water turned golden yellow. The Whattmann filter paper was used to filter the extract.

Preparation of ferric chloride solution

100 ml of 1mM Ferric chloride (FeCl₃) solution was prepared using distilled water.

Synthesis of iron oxide nanoparticle

In a beaker, a 1:1 mixture of the produced ferric chloride solution and *Solanum nigrum* leaf extract was added. The *Solanum nigrum* leaf extract is combined with ferric chloride solution to create a black, particle-filled solution. For three hours, a magnetic stirrer was used to continuously agitate the liquid. The generated solid product was extracted using centrifugation, cleaned three to four times with distilled water, and then allowed to air dry. After the dry product was calcined at 750°C for three hours, a powder with a reddish brown colour was produced. Karthikamurganatham *et al* (2015).

Sample Characterization

The synthesized Fe₂O₃ NPs were exposed to various characterization methods to identify their specific properties. For optical properties, UV-Vis spectrometer was used to record the absorption spectrum. SEM was used to determine the shape of Fe₂O₃ NPs. The functional groups were identified by recording the FTIR spectrum. And particle size of the Fe₂O₃ NPs were characterized by using XRD.

Antibacterial Activity

Bacteria strains

The Iron nanoparticle synthesis from the extracts (SN-NPs) were tested for antimicrobial activity against two Gram-positive bacterial strain and two Gram-negative-bacterial strain.

Agar well diffusion assay

Fe₂O₃ NPs tested for their antibiotic sensitivity pattern against different types of Gram-positive and negative pathogens, which are known to develop antibiotic resistance, such as *Staphylococcus aureus*, *Streptococcus pyogenes*, *Klebsiella pneumonia*, and *Escherichia coli*. Colonies from different types of bacteria were inoculated in L.B. broth and incubated for 24 hours at 37°C in an incubator. The bacterial cultures were then diluted to 1: 100 (equivalent to

a cfu of 10^8). A 50–100 μL of the bacteria was taken, and a streak was made on nutrient agar (L. B. agar) medium using a sterile spreader in all directions. The antibiotic disks were applied with aseptic precautions. The disks were soaked in different samples of iron oxide nanoparticles and extracted and placed with centers at least 30 mm apart. The plate was incubated at 37°C in an incubator for 24 hrs. After incubation, the zone of inhibition around the disks was observed and measure.

Results and Discussion

Visual Observation

The addition of Ferric Chloride(B) solution to the *Solanum nigrum* leaf extract(A) produces a brown colour solution(C) suspended with particles. Upon calcination at 750°C for three hours, brownish-black colour powder(D) was obtained.

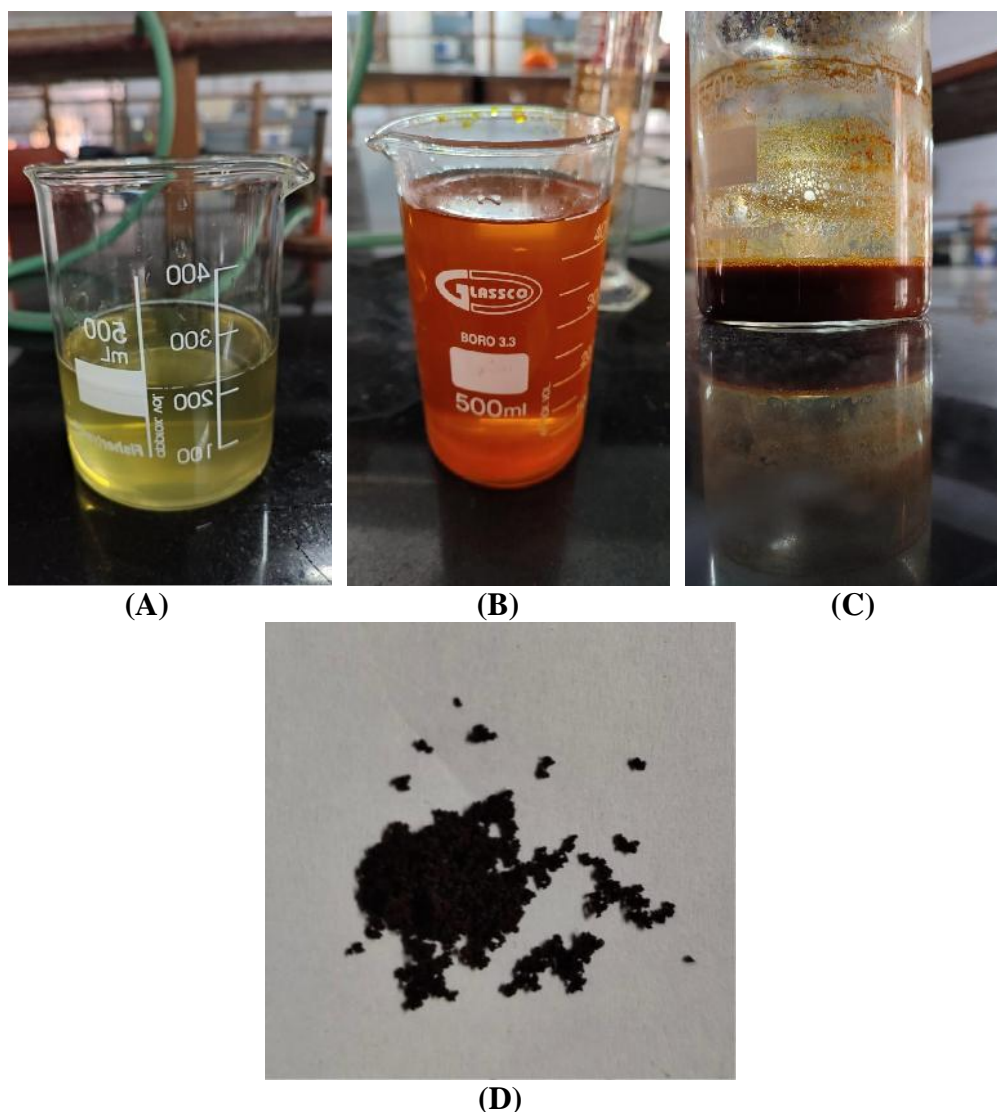


Fig 1: (A) *Solanum nigrum* leaf extract (yellow colour); (B) Ferric Chloride solution (reddish yellow colour); (C) Mixture of leaf extract and Ferric Chloride solution (brown colour);(D) Synthesized Iron Oxide compound (brownish black colour)

X Ray Diffraction (XRD)

Figure 2 shows the X-ray diffraction spectrum of the synthesized Fe₂O₃ NPs using the *Solanum nigrum* extract. The spectrum was recorded at a speed scan of 1 degree per minute in a 2theta/theta range of 10–70 degrees at an X-ray wavelength of 1.54 nm. The indexed diffraction peaks shown in the figure represent the crystalline phase of Fe₂O₃NPs. The match of the peaks with JCPDS card number 019-0629 for Fe₂O₃NPs confirms the synthesis of hematite -Fe₂O₃ nanocrystals. The average crystallite size D was calculated by the Debye Scherrer equation:

$$D = 0.9 / (\cos \theta)$$

Where,

is the wavelength of X-ray,

is the Braggs angle in radians,

is the full width at half maximum of the peak.

The crystallite size D was found to be an average size of 24.1 nm. The results of the XRD analysis supported the tetragonal structure of -Fe₂O₃ nanoparticles. By comparing the SEM and XRD size values, one concludes that there are multiple crystals in one particle in the case of single-crystal nanoparticles; the crystallite size and particle size are the same.

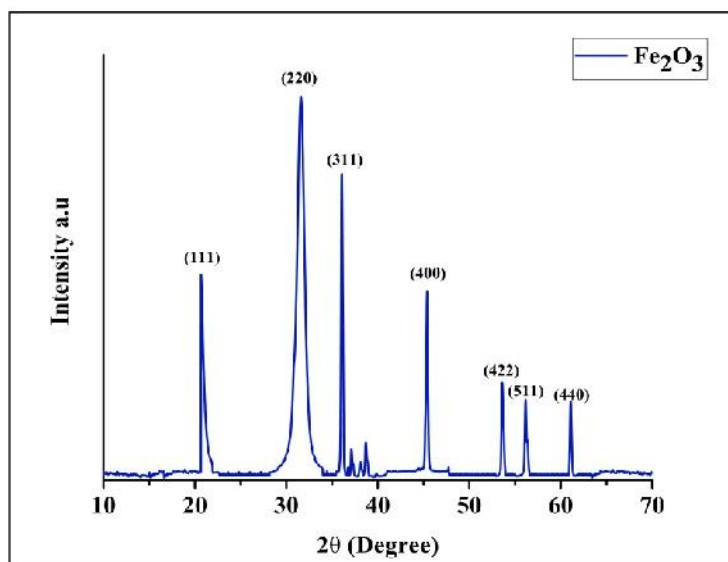


Fig 2: Xray Diffraction of Fe₂O₃ NPs synthesized using *Solanum nigrum* leaves extract.

Scanning electron microscopy (SEM) and energy dispersive X-ray spectroscopy (EDS)

SEM and EDS the morphologies of Fe₂O₃ NPs synthesized using *Solanum nigrum* leaves extract samples were examined using the SEM technique. The surface of the material was significantly magnified in the micrographs produced by scanning electron microscopy (SEM). Fig 3 (a) shows greater magnification micrographs of Fe₂O₃ NPs samples taken at different magnifications.

The synthesized Fe₂O₃ NPs had a uniform shape and a smooth surface, and this surface demonstrated the Hexagonal structural characterization of Fe₂O₃ NPs. Figure 3 (b) represents the EDS analysis showing firm Fe₂O peaks in Fe₂O₃ NPs, as shown in Table 1.

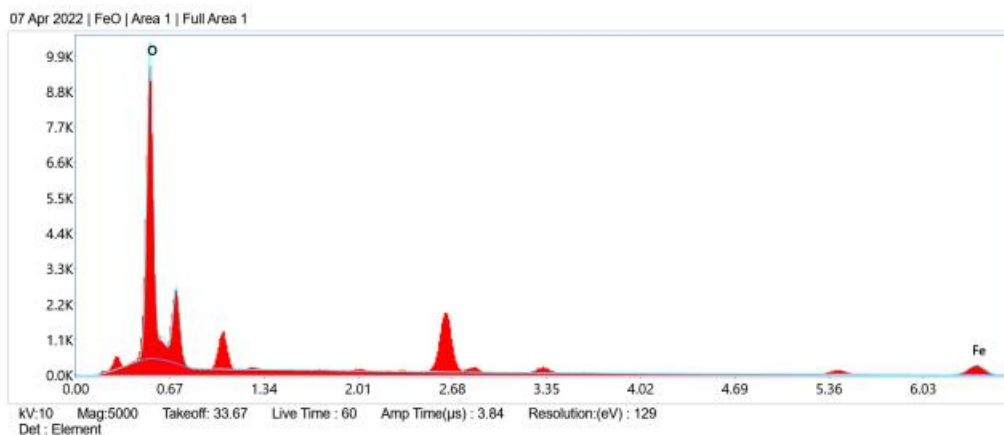
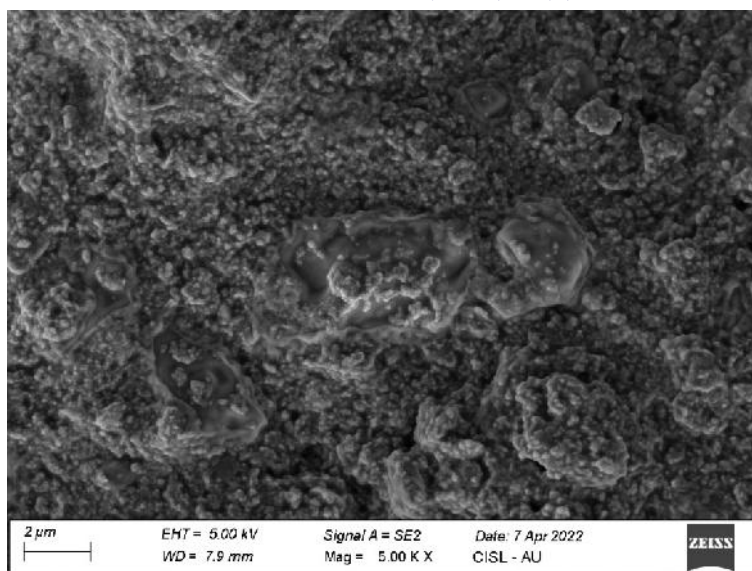


Fig 3 (a) SEM image and (b) EDS graph of Fe₂O₃ NPs synthesized using *Solanum nigrum* leaves extract.

Table 1: EDS measurements of Fe₂O₃ NPs synthesized using *Solanum nigrum* leaves extract.

Element	Weight %	Atomic %
O	45.27	74.28
Fe	54.73	25.72

UV-Vis spectroscopy

Ultraviolet-visible spectroscopy was used to monitor the degree of oxidation of Fe₂O₃ NPs. Fe₂O₃ NPs spectrum produced is illustrated in Fig 4. The absorption peak was discovered at 222 nm.

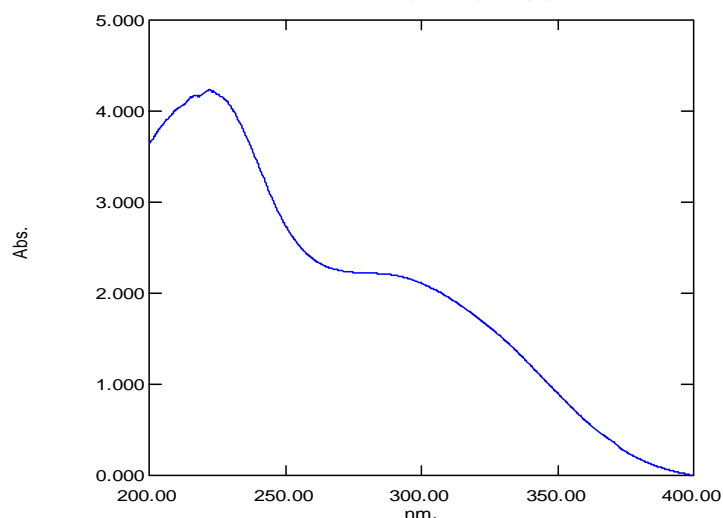


Fig 4: UV-Visible spectroscopy of Fe₂O₃ NPs synthesized using *Solanum nigrum* leaves extract.

Antibacterial activity

The antibacterial activity of Fe₂O₃ against bacterial pathogens was tested by a well diffusion method. The antibacterial potential of the biosynthesized Fe₂O₃ NPs using *Solanum nigrum* leaf extract is tested against *Escherichia coli*, *Staphylococcus aureus*, *Streptococcus pyogenes*, *Klebsiella pneumonia* and shown in Fig.5, 6 and table 2. The biosynthesized Fe₂O₃ NPs showed antibacterial effect on all the tested bacterial strains. The bactericidal effect of Fe₂O₃ NPs was found higher for Gram-negative bacteria than

Gram-positive bacteria and was based on the difference in the structural composition of Gram-positive and Gram-negative bacteria. *Solanum nigrum* leaf extract-derived Fe₂O₃ NPs showed the maximum zone of inhibition was observed against *Klebsiella pneumoniae* (24 mm) followed by *Escherichia coli* (20 mm), *Staphylococcus aureus* (18 mm) and *Streptococcus pyogenes* (16 mm) at 100 µl concentration. Similar results on antibacterial effect for *Klebsiella pneumonia* and *E. coli* by Fe₂O₃ NPs were reported previously in the literature.

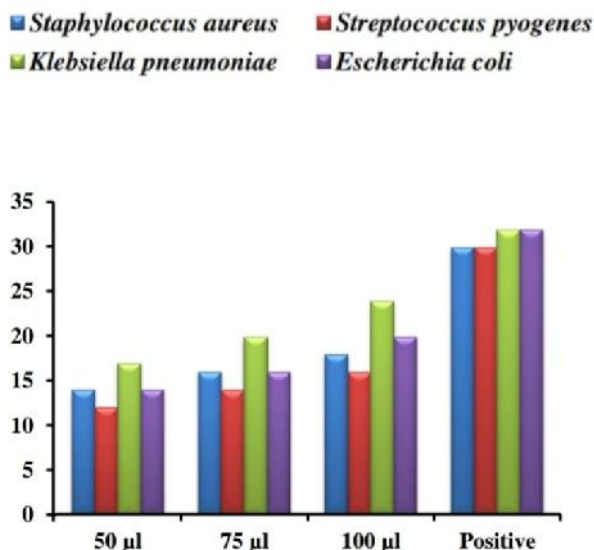


Fig 5: Graphical representation of zone of inhibition of Fe₂O₃ NPs synthesized using *Solanum nigrum* leaves extract.

The mechanism of antibacterial activity of Fe₂O₃ NPs may be attributed to the penetration and disintegration of the membrane by smaller sized NPs which lead to cell lysis. The release of H₂O₂ from the surface of Fe₂O₃ NPs also reported as the possible mechanism for bactericidal activity. The generation of H₂O₂ is highly depended on the surface area of Fe₂O₃ and the generated H₂O₂ penetrates the cell membrane and

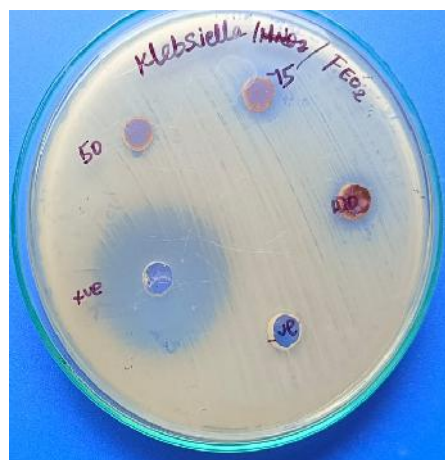
cause damage to kill the bacteria. Due to the presence of alkaloids, terpenoids, flavonoids, tannins, carbohydrates, sterols, saponins, proteins, and amino acids in *Solanum nigrum* leaf extract showed potential bioreducing activity and also bactericidal activity against the tested bacteria which could be useful for biomedical applications.

Table.2. Zone of inhibition of Fe₂O₃ NPs synthesized using *Solanum nigrum* leaves extract.

Bacterial pathogens	Zone of inhibition mm				
	50µl	75 µl	100 µl	Positive	Negative
<i>Staphylococcus aureus</i>	14	16	18	30	-
<i>Streptococcus pyogenes</i>	12	14	16	30	-
<i>Klebsiella pneumoniae</i>	17	20	24	32	-
<i>Escherichia coli</i>	14	16	20	32	-



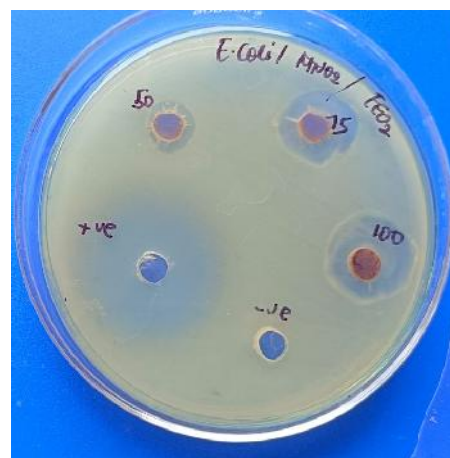
Streptococcus pyogenes



Klebsiella pneumoniae



Staphylococcus aureus



Escherichia coli

Fig 6: Antibacterial activity of Fe₂O₃ NPs synthesized using *Solanum nigrum* leaves extract.

Conclusion

Iron oxide nanoparticles were successfully synthesized by biogenic route using *Solanum nigrum* leaf extract. UV-Visible spectroscopy analysis confirmed the synthesis of Fe₂O₃ by the existence of band at 222 nm. SEM analysis revealed that the synthesized Fe₂O₃ were in Hexagonal structure. Crystal size (24.1) of the Fe₂O₃ was confirmed by XRD analysis. EDS explained that the Fe₂O₃ peaks are present in Fe₂O₃ NPs synthesized using *Solanum nigrum* leaves extract. Fe₂O₃ showed better antibacterial activity against Gram-negative bacteria than Gram-positive bacteria. Fe₂O₃ NPs synthesized *Solanum nigrum* leaf extract could be utilized as antibacterial agent in biomedical, textile, and food industries.

References

1. Albrecht MA, Evans CW, Raston CL. Green chemistry and the health implications of nanoparticles. *Green Chem* 2006; 8:417-32.
2. De D, Mandal MM, Gauri SS, Bhattacharya R, Ram S, Roy SK. Antibacterial effect of lanthanum calcium manganite (La_{0.67}Ca_{0.33}MnO₃) nanoparticles against *Pseudomonas aeruginosa* ATCC 27853. *J Biomed Nanotechnol* 2010; 6:138-44.
3. Dixon MB, Falconet C, Ho L, Christopher WK, O'Neill BK, Newcombe G. Removal of cyanobacterial metabolites by nano filtration from two treated waters. *J Hazard Mater* 2011; 1882:88-95.
4. Sastry M, Ahmad A, Khan MI, Kumar R. Microbial nanoparticle production. In: Niemeyer CM, Mirkin CA, editors. *Nanobiotechnology: Concepts, Applications and Perspectives*. Weinheim: Wiley-VCH Verlag GmbH and Co. KGaA; 2004. P. 126-35.
5. Bhattacharya D, Rajinder G. Nanotechnology and potential of microorganisms. *Crit Rev Biotechnol* 2005; 25:199-204.
6. Mohanpuria P, Rana NK, Yadav SK. Biosynthesis of nanoparticles: technological concepts and future applications. *J Nano Part Res* 2008; 10: 507-17.
7. Rosi NL, Giljohann DA, Thaxton, CS, Lytton-Jean AKR, Han MS, Mirkin CA. Oligonucleotide-modified gold nanoparticles for intracellular gene regulation. *Science* 2006; 312:1027.
8. Shchukin DG, Schattka JH, Antonietti M, Caruso RA. Synthesis of nano-sized magnetic ferrite particles inside hollow polyelectrolyte capsules. *J PhysChem B* 2003; 107:952.
9. Shobha G, Vinutha M, Ananda S. Biological synthesis of copper nanoparticles and its impact – a Review. *Int J Pharm Sci Invent* 2014; 38:28-38.
10. Sawai J. Quantitative evaluation of antibacterial activities of metallic oxide powders (ZnO, MgO and CaO) by conductimetric assay. *J Microbiol Methods* 2003; 54:177-82.
11. Shahverdi AR, Fakhimi A, Shahverdi HR, Minaian S. Synthesis and effects of silver nanoparticles on the antibacterial activity of different antibiotics against *Staphylococcus aureus* and *Escherichia coli*. *J Nanomedicine* 2007; 3: 168-17.
12. Pattanayak M, Nayak PL. Green synthesis and characterization of zero valent iron nanoparticles from the leaf extract of *Azadirachta indica* (Neem). *World J Nano Sci Tech* 2013; 2:6-9.
13. Sarkar B, Bhattacharjee S, Daware A, Tribedi P, Krishnan KK, Minhas PS. Selenium nanoparticles for stress resilient fish and livestock. *Nanoscale Res Lett* 2015;10:371.
14. Sathishkumar M, Sneha K, Won SW, Cho CW, Kim S, Yun YS. *Cinnamom zeylanicum* bark extract and powder mediated green synthesis of nano-crystalline silver particles and its bactericidal activity. *Colloids Surf B Biointerfaces* 2009; 73: 332-8.

15. Latha N, Gowri M. Bio synthesis and characterization of Fe₃O₄ nanoparticles using *Carica papaya* leaves extract. Int J Sci Res 2014; 3: 1551-6.
16. Kumar A, Singhal A. Synthesis of colloidal -Fe₂O₃ nanostructures-influence of addition of Co²⁺ on their morphology and magnetic behaviour. J Nanotech 2007; 18: 475703.
17. Lee J, Isobe T, Senna M. Preparation of ultrafine Fe₃O₄ particles by precipitation in the presence of PVA at high pH. J Colloid Interface Sci 1996; 177:490.
18. Duman O, Tunc S. Electrokinetic and rheological properties of Na-bentonite in some electrolyte solutions. Micropor Mesopor Mat 2009; 117: 331-8.

Access this Article in Online	
	Website: www.ijarbs.com
	Subject: Nanotechnology
Quick Response Code	
DOI: 10.22192/ijarbs.2022.09.08.013	

[How to cite this article:](#)

M. Rashmi Sulthana and M. Shenbagam Madhavan. (2022). Green synthesis, characterization, and antibacterial activity of iron oxide nanoparticles derived from *Solanum nigrum* leaf extract. Int. J. Adv. Res. Biol. Sci. 9(8): 131-139.

DOI: <http://dx.doi.org/10.22192/ijarbs.2022.09.08.013>