International Journal of Advanced Research in Biological Sciences ISSN: 2348-8069 www.ijarbs.com

(A Peer Reviewed, Referred, Indexed and Open Access Journal) DOI: 10.22192/ijarbs Coden: IJARQG (USA) Volume 10, Issue 2 -2023

Research Article

DOI: http://dx.doi.org/10.22192/ijarbs.2023.10.02.008

Evaluation of the microbial quality and safety of regulated and unregulated liquid herbal preparations in Benin City, Nigeria.

Omoruyi, Z^{1*}, Okundaye M. I.² and Egunjobi, T. O.³

 ^{1,2} Department of Medical Laboratory Science, School of Basic Medical Sciences, University of Benin, Benin City, Edo State, Nigeria.
 ³ Department of Medical Microbiology, School of Basic Clinical Sciences, College of Health Sciences, Igbinedion University Okada, Edo State, Nigeria. Correspondence: Dr. (Mrs) Z. Omoruyi
 Department of Medical Laboratory Science, School of Basic Medical Sciences, University of Benin.
 E-mail: zainab.omoruyi@uniben.edu

Abstract

Due to people's unhappiness with orthodox treatment, prior positive experiences, family traditions, and poverty, the use of herbal drugs has expanded globally. Unhygienic production conditions are typically one of the drawbacks of herbal products. The aim of this study was to evaluate the microbial quality and safety of regulated and unregulated liquid herbal preparations in Benin City, Nigeria. Standard microbiological procedures were employed in the identification of the isolated microbes and the evaluation of the total viable bacterial count in the 50 herbal preparations that were tested. A larger proportion (36%) of the unregulated herbal preparations was contaminated with bacteria as compared with regulated ones (4%). *Klebsiella pneumoniae* was the predominant bacterium isolated from unregulated herbal products. A proportion of 2% and 26% of the regulated and unregulated herbal preparations, respectively, exceeded the World Health Organization's acceptable limit for microbial contamination. In Benin City, there are a lot of unregulated herbal remedies with microbial contamination that is higher than the WHO's permitted level for aerobic and coliform bacteria. The regulatory agencies should intensify efforts to ensure that all herbal medicines marketed in the country are lawfully registered and produced in accordance with current Good Manufacturing Practice standards.

Keywords: Good Manufacturing Practice (GMP), Herbal preparation, Microbial contaminations, NAFDAC.



Introduction

According to the World Health Organization (WHO), traditional herbal medicine includes herbs, herbal preparations, herbal materials, and completed herbal products that include plant parts or other plant materials as active components and are used to cure or prevent a variety of illnesses (Tilburt and Kaptchuk, 2008). In low-and middleincome countries, about 80% of the population utilizes herbal drugs as their main source of primary health care (Ugbomoiko et al., 2022; Kretchy et al., 2021; Umairet al., 2017; WHO, 2002b). In many African nations, herbal preparation is the first option for treating children at home who have a high fever caused by malaria (Aschwanden, 2001). These products are reliable, simple to use, accessible, affordable, and believed to be safer than orthodox medicine (Abualhasan et al., 2020; Farrington et al., 2019). Due to the region's rich biodiversity, there is currently a trend in Nigeria toward employing a variety of herbal preparations as alternative medicines. Traditional herbal healers in Nigeria cure a wide range of illnesses with different herbal remedies, including skin conditions, cough, seizures, neonatal fistula, diarrhea, etc. (Sofowora, 1982). The majority of these herbal products are used as concoctions, which are soups or drinks that are often created by boiling materials, or as infusions, which involve soaking plant material and letting it stand for varying amounts of time (Adeleye and Opiah, 2003).

In the developing world, the microbial safety of herbal medicines is a big concern because there are worries that they may be prepared in unhygienic ways and put consumers in danger (Rajkumar and Sriram, 2021; Abtahi and Nourani, 2017; Khattak, 2012). These risks include those related to the nervous system, the heart, and the blood (Palmer *et al.*, 2003). Microbial pollutants that produce toxins are frequently to blame for these negative impacts. As a result, it is crucial to recognize the microbiological contaminants in herbal preparations as signs of their quality and safety (Schweiggert *et al.*, 2005).Herbal goods may become contaminated by microorganisms at any point in the production process, from planting and harvesting to packaging, shipping, and storage (Kosalec et al., 2009). The packaging for these goods is frequently poor as well; the finished preparations are frequently placed in unlabeled recycled plastic bottles (Igbeneghu and Lamikanra, 2016). Staphylococcus aureus. Salmonella typhi, Escherichia coli, Enterobacter spp., Bacillus spp., Klebsiella pneumoniae, Proteus mirabilis, and Pseudomonas aeruginosa are a few of the prevalent bacteria detected in herbal medicines (de Sousa et al., 2020; Yesuf et al., 2016).

Because of this, pharmaceutical businesses are expected to follow the guidelines of Good Manufacturing Practice (FDA, 2015), and their products must be subjected to strict quality controls. Additionally, the entire medication manufacturing process must pass quality assurance tests at every stage (Igbeneghu and Lamikanra, 2016). However, it must be acknowledged that only the large pharmaceutical firms have the resources to follow the guidelines of modern GMP (Igbeneghu and Lamikanra, 2016). Small pharmaceutical companies many of which are located in nations with struggling economies are unable to finance machinery or the hiring of qualified personnel to ensure that their goods are of consistently high quality (Okeke and Lamikanra, 2001). Nigeria's situation is made worse by the existence of a sizable informal sector, which is in charge of the small-scale production of numerous unregistered and typically unstandardized medications using crude equipment and raw plant-derived materials that are extremely vulnerable extensive to contamination (Igbeneghu and Lamikanra, 2016). The relevant regulatory organizations have no unregistered control over these herbal However. medications. they cannot be disregarded because they are widely accessible, particularly in rural areas where there is a lack of modern pharmaceutical coverage (Igbeneghu and Lamikanra, 2016). Therefore, this study aimed to evaluate the microbial quality and safety of regulated and unregulated liquid herbal preparations in Benin City, Nigeria.

Method

Study Area and Design

This cross-sectional study was carried out in Benin City. The Benin City is the capital of Edo State, located in the South-South region of Nigeria (latitude $6^{\circ}20\ 00$ " N and longitude $5^{\circ}37'20$ "E). Benin City has an estimated area of 1,204 km² and a population of over 1,500,000 according to the 2006 census (Encyclopedia Britannica, 2022).

Sample Collection and Preparation

Ten (10) regulated and forty (40) unregulated liquid herbal preparations were randomly purchased from 3 major areas across Benin City, Nigeria; from hawkers, herbal shops, and drug stores. These areas were Adolor, Technical, and Uselu. Details regarding the dosage and uses of each product were collected from the vendors and recorded. To avoid bias, the reason for purchasing the preparations was not made known to the vendors. All samples were transported in a sterile screw-capped bottle to the Medical Microbiology Laboratory of the University of Benin Teaching Hospital (UBTH) in a cold box within one hour of collection. The preparations were stored at 4 °C until use.

Laboratory Analysis

Identification of isolated bacterial species

Ten millilitres of the samples were centrifuged for 5 minutes at 5000 rpm. The pellets were then inoculated onto MacConkey agar, Blood agar, Eosin Methylene Blue agar, and Deoxycholate Citrate agar. For 18 to 24 hours, the culture plates were incubated at 37°C. Isolated bacteria were identified by standard microbiological procedures (Cheesbrough, 2006).

Evaluation of total viable bacterial count

The samples were mixed vigorously to homogenize and then serially diluted by ten-folds

up to 10^{-5} . The microbiological analysis was performed in triplicate. One millilitre of each dilution was pipetted into each Nutrient Agar plate with a sterile pipette and incubated for 24 hours at 37°C. After the incubation period, the average number of colonies was multiplied by the dilution factor to determine the colony-forming units per milliliter (CFU/ml). The CFU/ml of the sample that was obtained was compared to WHO standards. According to the WHO criteria for aerobic bacteria, herbal preparations with > 10^5 CFU/ml were considered unsatisfactory (WHO, 2007a).

Antimicrobial susceptibility test

Isolates were cultivated overnight on nutrient agar and emulsified in 3ml of sterile physiological saline to achieve turbidity corresponding to 0.5 McFarland standards. Using a sterile, non-toxic cotton swab, the Mueller-Hinton agar plates were inoculated with the standardized inoculum. The selected antibiotic discs (Cefoxitin [30 µg], Piperacillin [100 µg], Gentamycin [10 µg], and Meropenem [10 µg]) by Thermo ScientificTM, MA, USA; were placed on the agar plates and incubated at 37°C for 24 hours.

Statistical Analysis

Descriptive statistical analysis was carried out using SPSS (Statistical Package for the Social Sciences) Version 23 Software. The Chi-square test was employed to determine whether category variables were related. For all statistical tests, a P value less than 0.05 was considered significant.

Results

Characteristics and intended use of herbal preparations sold in Benin City.

Among the 50 herbal preparations tested, 15 (30%) were purchased each from the Adolor and Technical areas of Benin City, while 20 (40%) were from the Uselu area of the city. Forty (80%) of the herbal preparations were unregistered and unregulated by the appropriate Regulatory body. Thirty-nine (78%) of the herbal products were

prepared with water (Table 1). Each of the herbal preparations tested in this study has one or more therapeutic purposes according to their sellers (Table 2).

Table 1: Characteristics of herbal preparations sold in Benin City, Nigeria.

| Herbal Preparation | Frequency (%) | |
|--------------------|---------------|--|
| Regulation Status | | |
| Regulated | 10 (20.0) | |
| Unregulated | 40 (80.0) | |
| Total | 50 (100.0) | |
| Solvent | | |
| Water | 39 (78.0) | |
| Alcohol | 11 (22.0) | |
| Total | 50 (100.0) | |
| Area | | |
| Adolor | 15 (30.0) | |
| Technical | 15 (30.0) | |
| Uselu | 20 (40.0) | |
| Total | 50 (100.0) | |

Table 2: Intended use of herbal preparations sold in Benin City, Nigeria.

| | Herbal Preparation | | |
|--------------------------|--------------------|-------------|--|
| Purpose | Regulated | Unregulated | |
| | Ň (%) | N (%) | |
| Asthma | - | 1 (2.5) | |
| Blood tonic | - | 1 (2.5) | |
| Body pain | - | 3 (7.5) | |
| Cough/Pharyngitis | 1 (10.0) | 3 (7.5) | |
| Deworming | - | 1 (2.5) | |
| Diarrhoea/Dysentery/Pile | - | 4 (10.0) | |
| Immune booster | 2 (20.0) | - | |
| Malaria | - | 2 (5.0) | |
| Multipurpose | - | 2 (5.0) | |
| Reproductive health | 5 (50.0) | 3 (7.5) | |
| Rheumatism/Arthritis | 2 (20.0) | 4 (10.0) | |
| Skin infections | - | 1 (2.5) | |
| STIs/Gonorrhoeae | - | 1 (2.5) | |
| Stomach pain | - | 5 (12.5) | |
| Stomach ulcer | - | 2 (5.0) | |
| Footh infections | - | 1 (2.5) | |
| Гурhoid | - | 3 (7.5) | |
| UTI | - | 3 (7.5) | |
| Total | 10 (100.0) | 40 (100.0) | |

Prevalence and total viable counts of isolated bacteria species

Overall, 20 (40%) of the tested herbal preparations were contaminated with bacteria, out of which 36% were from unregulated herbal products. A total prevalence of 4% was observed each for *S. aureus*, *E. coli*, and Enterobacter spp

among the tested herbal products. However, *K. pneumoniae* had a prevalence of 10%, followed by *K. oxytoca* and *P. mirabilis* with 8% each (Table 3). The microbial counts of isolated bacteria ranged from 2.8 x 10^4 CFU/ml to 3.1 x 10^4 CFU/ml and 3.8 x 10^4 CFU/ml to 12.6 x 10^8 CFU/ml for the regulated and unregulated herbal products, respectively (Data not shown).

Table 3: Bacterial species isolated from regulated and unregulated herbal preparations.

| De starie Iselete | | Herbal Preparation | | |
|-------------------|-----------------|--------------------|-------------------------|--|
| Bacteria Isolate | Regulated N (%) | Unregulated N (%) | N (%) Total N (% | |
| S. aureus | 1 (2.0) | 1 (2.0) | 2 (4.0) | |
| E. coli | 1 (2.0) | 1 (2.0) | 2 (4.0) | |
| K. pneumoniae | - | 5 (10.0) | 5 (10.0) | |
| K. oxytoca | - | 4 (8.0) | 4 (8.0) | |
| P. mirabilis | - | 4 (8.0) | 4 (8.0) | |
| Enterobacter spp | - | 2 (4.0) | 2 (4.0) | |
| Citrobacter spp | 1 (2.0) | 1 (2.0) | 1 (2.0) | |
| Total | 2 (4.0) | 18 (36.0) | 20 (40.0) | |

Microbial quality of the herbal preparations

In general, 2% and 26% respectively, of the regulated and unregulated herbal preparations exceeded the acceptable limit of microbial contamination according to the WHO standards (WHO, 2007a). Although not statistically

significant, herbal products prepared with water were more contaminated with bacteria than those prepared with alcohol. Similarly, herbal products that are unregulated and prepared with water are 3 times more likely to exceed the acceptable microbial limit compare to those that are regulated and prepared with alcohol (Table 4).

Table 4: Microbial quality of herbal preparations in Benin City, Nigeria.

| Herbal Products | Microbial Quality | | | |
|----------------------|---------------------|-----------------------|-----------------------|---------|
| | Acceptable N (%) | Unacceptable N (%) | Odd Ratio (95% CI) | P-value |
| Regulation Status | | | | |
| Regulated | 9 (18.0) | 1 (2.0) | 1 | |
| Unregulated | 27 (54.0) | 13 (26.0) | 4.33 (0.49-37.93) | 0.1852 |
| Total | 36 (72.0) | 14 (28.0) | | |
| Solvent | | | | |
| Water | 26 (52.0) | 13 (26.0) | 5.00 (0.58-43.39) | 0.1443 |
| Alcohol | 10 (20.0) | 1 (2.0) | 1 | |
| Total | 36 (72.0) | 14 (28.0) | | |
| Area | | | | |
| Adolor | 12 (24.0) | 3 (26.0) | 0.69 (0.12-3.79) | 0.6668 |
| Technical | 11 (20.0) | 4 (2.0) | 1 | |
| Uselu | 13 (20.0) | 7 (2.0) | 1.48 (0.34-6.42) | 0.6001 |
| Total | 36 (72.0) | 14 (28.0) | | |
| | × / | 63 | | |

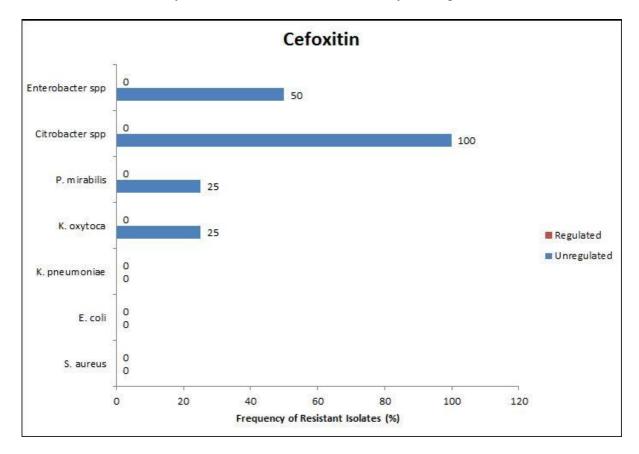
| Regulation Status and Solvent | | | | |
|-------------------------------------|-----------|-----------|-------------------|--------|
| Unregulated and water based | 23 (46.0) | 13 (26.0) | 3.39 (0.37-31.34) | 0.2817 |
| Unregulated and alcohol based | 4 (8.0) | - | 0.48(0.02-14.70) | 0.6752 |
| Regulated and water based | 3 (6.0) | - | 0.62 (0.02-19.59) | 0.7855 |
| Regulated and alcohol based | 6 (12.0) | 1 (2.0) | 1 | |
| Total | 36 (72.0) | 14 (28.0) | | |

Acceptable limit: Aerobic bacteria – 10⁵ CFU/ml, *Escherichia coli* – 10 CFU/ml(WHO, 2007a).

Resistance of isolated bacterial species to antibiotics

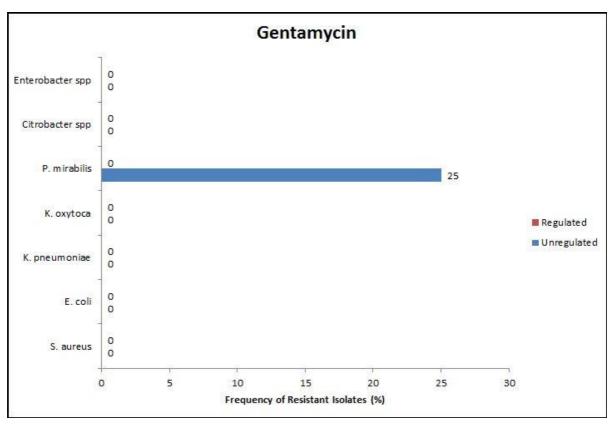
All the bacterial species isolated from the regulated herbal preparations were susceptible to all tested antimicrobials. Only 1 out of the 2

Enterobacter spp isolated from the unregulated herbal products showed resistance to Cefoxitin. However, both isolates were resistant to Piperacillin. Furthermore, 25% of the *P. mirabilis* isolated showed resistance to Cefoxitin and Gentamycin (Figure 1a-d).

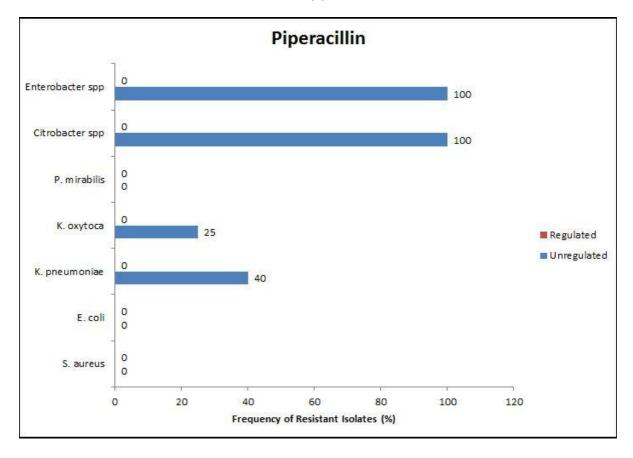


(a)



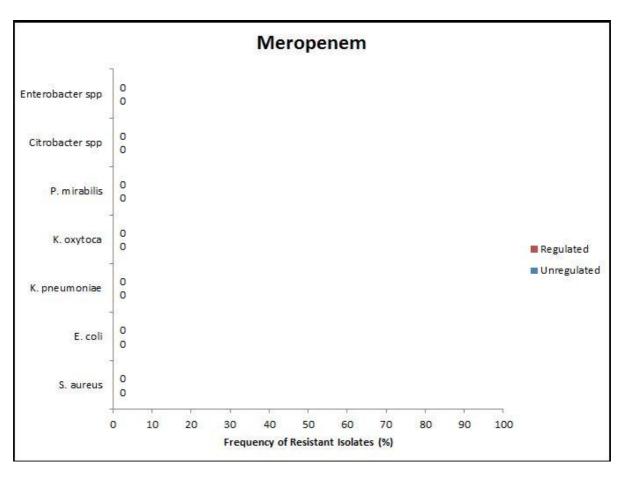


(b)



(c) 65

Int. J. Adv. Res. Biol. Sci. (2023). 10(2): 59-71



(d)

Figure 1(a-d): Prevalence of bacteria isolates resistant to: (a) Cefoxitin, (b) Gentamycin, (c) Piperacillin, and (d) Meropenem.

Discussion

Although herbal products are becoming more and more popular around the world, one barrier to their acceptability is the absence of a uniform quality control profile (Kunle et al., 2012). Lack of systems to track herbal medicine contaminants like microbes and chemicals, as well as providers' judgment of best practices, could be harmful health risk factors for herbal medicine users (WHO, 2019). These contaminations of herbal preparations both lessen their efficiency and put the health of their users in grave danger (Turkson et al., 2020). In the present study, 80% of the preparations were unregistered herbal and unregulated. There is a high proliferation of unregulated herbal products in many developing countries such as Nigeria owing to their cheaper cost and readily available than the regulated ones.

Unregulated herbal preparations are sold by street hawkers and in many unlicensed herbal shops in Benin City, Nigeria. Poverty and ignorance of the health risks associated with the use of unregulated herbal products are major reasons for the general acceptance of these products. According to the World Health Organization, limited financial resources are a key reason people utilize herbal medicine (WHO, 2011).

Unhygienic production conditions are typically one of the drawbacks of herbal products (Nwoko and Mgbeahuruike, 2011; Oyetayo, 2008). Their negative consequences range in severity, including deaths (Justin-Temu *et al.*, 2009). A prevalence of 40% microbial contamination was observed for the regulated and unregulated herbal preparations, out of which 36% was from the unregulated herbal products. Several studies had reported higher rates of bacterial contamination than what was observed in this study (Darkwah et al., 2022; Kira et al., 2021; Kalumbi et al., 2020; Igbeneghu and Lamikanra, 2016). This disparity may be due to the fact that some of these studies were conducted in countries with no regulation governing the production of herbal products resulting in the proliferation of low-quality herbal products (Kira et al., 2021; Kalumbi et al., 2020). The primary duty of regulatory agencies is to make sure that patients receive medication that is guaranteed to be pure, safe, potent, and effective (Kunle et al., 2012). In Nigeria, the National Agency for Food and Drug Administration and Control (NAFDAC) is the agency responsible for regulating and controlling both local and imported herbal medicines in the country (Osuide, 2002). On the other hand, this result is in tandem with the reports of Abualhasan et al (2020) who reported a high rate of microbial contamination among unregulated herbal tea in Palestine. The high rate of microbial contamination observed among the unregulated herbal preparations may be due to the lack of resources to follow the guidelines of current Good Manufacturing Practices. Unregulated herbal products in Nigeria are mainly homemade, produced by individuals that are unable to purchase machinery that could ensure the high quality of their products (Okeke and Lamikanra, 2001).

The presence of Staphylococcus aureusin this study is lower than 8% and 49.2% reported in Tanzania (Kira et al., 2021) and Brazil (de Sousa et al., 2020), respectively. This difference in data may be due to the improvement in the packaging of herbal products over the years in Nigeria (Igbeneghu and Lamikanra, 2016). However, a lower prevalence of this pathogen had been reported in a previous study in Uganda (Walker et al., 2021). In our study, Citrobacterspp was isolated in 2% of the unregulated herbal products. This result is in accordance with the reports of Igbeneghu and Lamikanra (2016), but lower compared to the 30% reported in Blantyre, Malawi (Kalumbi et al., 2020). Citrobacter spp and S. aureus have been linked to food poisoning in humans (Brooks et al., 2013; Bai et al., 2012). The presence of Escherichia coli is an indication of fecal contamination (Edberg et al., 2000). This bacterium was isolated from 4% of the regulated unregulated herbal preparations. and This microorganism could be acquired by using contaminated water and containers to prepare herbal products. Utilizing plant parts that have been exposed to manure but have not been properly washed or disinfected is another possible source. Vegetables and other plant components have been documented to act as reservoirs for a variety of bacteria (Holden et al., 2009). Escherichia coli contamination has also been linked to poor harvesting and production methods (WHO, 2007b).

The rate of unregulated herbal preparations exceeding the permissible limit for microbial contamination in the present study (26%) is lower than 39.4% and 48% reported in Bangladesh (Nur et al., 2018) and Iran(Ameri et al., 2020), respectively. The high number of herbal products that fail the microbial test in these studies could be linked to the poor quality of water in some of these countries (Parvin et al., 2022; Fanack Water, 2021). Furthermore, in our study, herbal products prepared with water were observed to be more frequently contaminated than products prepared with alcohol. This is made worse when the product is unregulated. The use of medicinal herbal medications takes many various forms, and the manipulation and processing steps have a significant impact on the microbial quality of the finished preparation (Kneifel et al., 2002). Since boiling water is predicted to significantly reduce the viable counts by several log units and inactivate potential pathogens, the use of hot water extraction typically compensates for microbiological contaminations (Kneifel et al., 2002). In contrast, herbal preparations undergoing cold water extraction may include a significant number of microorganisms, and the ambient temperature extraction process typically promotes microbial multiplication. In general, ethanolextracted herbal tinctures offer hygienic conditions, however, the outcome will vary depending on the concentration of alcohol used (Kneifel et al., 2002).

Int. J. Adv. Res. Biol. Sci. (2023). 10(2): 59-71

Some herbal products may pose a major health risk to users due to contamination with bacteria carrying genes for antibiotic resistance (Kira et al., 2021). Enterobacterspp isolated in this study showed resistance to Cefoxitin and Piperacillin. Similarly, 25% of the Proteus mirabilis isolated were resistant to Cefoxitin and Gentamycin. A similar incidence of bacterial resistance to widely used antimicrobials was documented in studies conducted elsewhere (Kira et al., 2021). This observation may be the result of the misuse of antibiotics in animals and human health practices for the prevention and management of bacterial illnesses (Rahimi and Nayebpour, 2012; Esimone et al., 2007; Foster, 1983). Additionally, this resistance could also result from the overuse of herbs with antimicrobial properties, which causes bacteria to become resistant to antibiotics with similar chemical structures (Kalumbi, 2019; Kalumbi, 2018; Tiwari and Tiwari, 2011).

Conclusions

There are a substantial number of unregulated herbal products in Benin City with microbial contamination exceeding the WHO acceptable limit for aerobic and coliform bacteria. Alcoholbased herbal preparations are more microbially stable than those prepared with water. The regulatory authorities should intensify efforts to see that all herbal medicines in the country are duly registered and their productions follow the guidelines of current GMP. This can be achieved by recognizing small-scale herbal product vendors as alternative health care providers in the country and providing appropriate training and financial support to them. In addition, public enlightenment focused on the health hazards associated with the use of adulterated and unregulated herbal products should be put in place and emphasized.

Conflict of Interest Statement: None

References

Abtahi, F. and Nourani, S.L. 2017. The most important fungal diseases associated with some useful medicinal plants. In: Ghorbanpour, M., Varma, A. eds. Medicinal Plants and Environmental Challenges. Springer International Publishing, AG, New York, USA.279-293.

Abualhasan, M.N., Jaradat, N., Hawash, M., Khayat, R., Khatatbeh, E., Ehmidan, M. and Al-Atrash, M. 2020.Evaluation of Heavy Metal and Microbial Contamination in Green Tea and Herbal Tea Used for Weight Loss in the Palestinian Market. Evid Based Complementary Altern Med. 2020:763-770.

https://doi.org/10.1155/2020/7631562

- Adeleye, I.A. and Opiah, L. 2003. Antimicrobial activity of extracts of local cough mixtures on upper respiratory tract bacterial pathogens. West Indian Med J. 52:188-90.
- Ameri, A., Ekhtelat, M. and Shamsaei, S. 2020.Microbial indices of industrial and traditional medicinal herbs in Ahvaz, Iran.Foods Raw Mater. 8(1):134–139. DOI:http://doi.org/10.21603/2308-4057-2020-1-134-139.
- Aschwanden, C. 2001. Bulletin of the World Health Organization. 79(7):691-692.
- Bai, L., Xia, S., Lan, R., Liu, L., Ye, C. and Wang, Y. 2012. Isolation and characterization of cytotoxic, aggregative *Citrobacter freundii*. Plos ONE 7(3):e33054. https://doi.org/10.1371/journal.pone.0033 054
- Brooks, G.F., Carroll, K.C., Butel, J.S., Morse,
 S.A. and Mietzner, T.A. 2013.Jawetz,
 Melnick, & Adelberg's Medical
 Microbiology, 26thedn. McGraw-Hill
 Companies, Inc. New York, United State.
 p199-200.
- Cheesbrough M. 2006. District Laboratory Practice in Tropical Countries, Part 2.Second Edition. Cambridge University Press, New York, USA. p62-70.
- Darkwah, S., Agbettor, D., Codjoe F. and Donkor, E.S. 2022.Microbial Contamination of Herbal Preparations on the Ghanaian Market, Accra. Microbiol Insights. 15:1– 4.

https://doi.org/10.1177/117863612211396 0

- de Sousa, L.C.M., Fujishima, M.A.T., de Paula, L.B., Mastroianni, P.C., de Sousa, F.F.O. and da Silva, J.O. 2020. Microbial contamination in herbal medicines: a serious health hazard to elderly consumers. BMC Complement Med Ther. 20:17.
- Edberg, S.C., Rice, E.W., Karlin, R.J. and Allen, M.J. 2000.*Escherichia coli*: the best biological drinking water indicator for public health protection. J. Appl. Microbiol. 88:106-116.
- EMEA. 2005. Guidelines on Quality of Herbal Medicinal Products/Traditional Medicinal Products, EMEA/CVMP/814OO Review. European Agency for the Evaluation of Medicinal Products (EMEA), London.
- Encyclopedia Britannica. 2022. Benin City History & Facts. Retrieved 14 February 2023.
- Esimone, C.O., Oleghe, P.O., Ibezim, E.C., Okeh, C.O. and Iroha, I.R. 2007. Susceptibilityresistance profile of micro-organisms isolated from herbal medicine products sold in Nigeria. Afr. J. Biotechnol. 6(24):2766-2775. https://doi.org/10.5897/AJB2007.000-2442.
- Fanack Water. 2021. Water Resources and Quality in Iran. Available at : https://water.fanack.com/iran/waterresources-in-iran/#_ftn19. Accessed on 15 February 2023.
- Farrington, R., Musgrave, I.F. and Byard, R.W. 2019. Evidence for the efficacy and safety of herbal weight loss preparations. J Integr Med. 17:87–92.
- FDA. 2015. Facts about the Current Good Manufacturing Practices (CGMPs). Available at: http://www.fda.gov/Drugs/DevelopmentA pprovalProcess/Manufacturing/ucm16910 5.htm. Accessed on 14th February, 2023.
- Foster, T.J. 1983. Plasmid-determined resistance to antimicrobial drugs and toxic metal ions in bacteria. Microbiol.Rev. 47:361-409. https://doi.org/10.1128/mr.47.3.361-409.1983

- Holden, N., Pritchard, L. and Toth, L. 2009. Colonization out with the colon: plants as an alternative environmental reservoir for human pathogenic Enterobacteria. FEMS Microbiol.Rev. 33:689-703.
- Igbeneghu, O.A. and Lamikanra, A. 2016. Assessment of the microbial quality of some oral liquid herbal medicines marketed in Ile-Ife, Southwestern Nigeria. Afr. J. Microbiol. Res. 10(38):1618-1624. Doi:10.5897/AJMR2016.8237
- Justin-Temu, M. Lyamuya, E.F. and Makwaya, C.K. 2009. Sources of microbial contamination of local herbal medicines sold on the open market in Dar es Salaam, Tanzania. East Cent. Afr. J. Pharm. Sci.12:19-20.
- Kalumbi, M. 2018. Effect of processing method and concentration of *Mangifera indica* leaf extract on their antibacterial activity against *Staphylococcus aureus*. Int J Herb Med.; 6(6):116-119.
- Kalumbi, M. 2019. Effect of Vancomycin, tetracycline, *Persia americana* leaf extract and combinations thereof on antibacterial activity against pathogenic organisms. J PharmacognPhytochem. 8(1):2733-2737.
- Kalumbi, M.H., Likongwe, M.C., Mponda, J., Zimba, B.L., Phiri, O., Lipenga, T., Mguntha, T. and Kumphanda, J. 2020.
 Bacterial and heavy metal contamination in selected commonly sold herbal medicine in Blantyre, Malawi. Malawi Med J. 32(3):153-159.
- Khattak,F. 2012. Microbiological quality assessment of commercially available medicinal plants in Peshawar city, Pakistan. Pak J Bot. 44:1203-1208.
- Kira, J.D., Mkupasi, E.M., Katakweba, A.A.S. and Ngowi, H.A. 2021.Assessment of Bacterial Contamination in Herbal Medicine Products Vended in Morogoro Municipality, Tanzania. East Cent. Afr. J. Pharm. Sci. 24:21-28.
- Kneifel, W., Czech, E. and Kopp, B. 2002.Microbial contamination of medicinal plants – A review.Planta Med. 68:5-15.

Int. J. Adv. Res. Biol. Sci. (2023). 10(2): 59-71

- Kosalec, I., Cvek, J. and Tomic, S. 2009.Contaminants of medicinal herbs and herbal products, ArhHigRadaToksikol. 60:485–501.
- Kretchy, I.A., Koduah, A. and Opuni, K.F.M. 2021. Prevalence, patterns and beliefs about the use of herbal medicinal products in Ghana: a multicentre community based cross sectional study. Trop Med Int Health. 26:410-420.
- Kunle, O.F., Egharevba, H.O. and Ahmadu, P.O. 2012. Standardization of herbal medicines-A review. Int. J. Biodvers. Conserv. 4(3):101-112.
- Nur, F., Libra, U.K., Rowsan, P., Azad A.K. and Begum K. 2018. Assessment of Bacterial Contamination of Dried Herbs and Spices Collected from Street Markets in Dhaka. Bangladesh Pharma J. 21(2):96-100.
- Nwoko O.C. and Mgbeahuruike, L. 2011. Heavy metal contamination of ready-to-use herbal remedies in southeastern Nigeria. Pakistan Journal of Nutrition. 10(10):959– 964.
- Okeke. I.N. and Lamikanra, A. 2001. skin Bacteriological quality of moisturizing creams and lotions distributed in a tropical developing country.J.Appl. Microbiol. 91(5):922-928.
- Osuide, G. E. 2002. Chapter21 Regulation of herbal medicines in Nigeria: the role of the National Agency for Food and Drug Administration and Control (NAFDAC). AdvPhytomedicine. 1:249-258. DOI:10.1016/S1572-557X(02)80030-7
- Oyetayo, V.O. 2008.Microbial load and antimicrobial property of two Nigerian herbal remedies.Afr J Tradit Complement Altern Med. 5(1):74–78.
- Palmer, M.E., Haller, C., Mckinney, P.E., Klein-Schwartz, W., Tschirgi, A., Smolinske, S.C., Woolf, A., Sprague, B.M., Bartlett, W.D., Landzberg, B.R. 2003. Adverse events associated with dietary supplements: an observational study. Lancet. 361:101-106.
- Parvin, F., Haque, M. and Tareq, S.M. 2022. Recent status of water quality in Bangladesh: A systematic review, meta-

analysis and health risk assessment. Environ Challenges. 6:100-106. https://doi.org/10.1016/j.envc.2021.10041 6

Rahimi, E. and Nayebpour, F.J. 2012. Antimicrobial resistance of *Escherichia coli* O157:H7/NM isolated from feaces of ruminant animals in Iran. Cell Anim. Biol. 6:104-108.

https://doi.org/10.5897/JCAB11.082

- Rajkumar, D.B. and Sriram, P. 2021. Evaluation of microbial load of herbal raw materials: a necessary quality control measure to ensure safety of finished herbal preparations. Adv Biotech & Micro.16:555934.
- Schweiggert, U., Mix, K., Schieber, A. and Carle, R. 2005. An innovative process for the production of spices through immediate thermal treatment of the plant material. Innov Food Sci. 6:143-153.
- Sofowora, A. 1982.Medicinal plants and traditional medicine in Africa. Chichester: Wiley. p256.
- Tilburt, J.C. and Kaptchuk, T.J. 2008. Bull. World Health Organization. 86:594–599.
- Tiwari, R. and Tiwari, G. 2011. Use of antibiotics: from proceeding to contemporary. Scholars' Res J. 1:59–68.
- Turkson, B.K., Mensah, M.L.K., Sam, G.H., Mensah, A.Y., Amponsah, I.K., Ekuadzi, E., Komlaga, G. and Achaab, E. 2020.Evaluation of the Microbial Load and Heavy Metal Content of Two Polyherbal Antimalarial Products on the Ghanaian Market.Evid Based Complementary Altern Med. 2020:1014-1019.

https://doi.org/10.1155/2020/1014273

Ugbomoiko, D.O., Egunjobi, T.O., Omosigho, P.O., Olley, M., Osaiyuwu, C., Asemota.
P.A. and Omoruyi, Z. 2022. Antibacterial Activity of the Leaf and Root Extracts of *Sansevieria zeylanica* Against Strains of Methicillin-Sensitive and Resistant *Staphylococcus aureus*. Int. J Curr. Res. Med. Sci.; 8(11):1. http://dx.doi.org/10.22192/ijcrms.2022. 08.11.001

- Umair, M., Altaf, M. and Abbasi, A.M. 2017.An ethnobotanical survey of indigenous medicinal Plants in Hafizabad district, Punjab-Pakistan.PLoSOne. 12:e0177912.
- Walker, G., Musinguzi, B., Musa, K., Mutesi, C., Zamarano, H., Manirakiza, G., Muhwezi, I., Kassaza, K., Tusingwire, F., Opendi, C. and Itabangi. H. 2021. Assessment of Pathogenic Contamination and Antimicrobial Activity of Selected Herbal Medicinal Remedies in Mbarara City, South Western Uganda. Research Square. 1-22. https://doi.org/10.21203/rs.3.rs-582967/v
- World Health Organization.2002a. General Guidelines for Methodologies on Research and Evaluation of Traditional Medicine. World Health Organization, Geneva.
- World Health Organization. 2002b. Traditional Medicine Strategy 2002–2005. World Health Organization.

- World Health Organization.2007a. Guidelines for assessing quality of herbal medicines with reference to contaminants and residues. Geneva: World Health Organization. p105.
- World Health Organization.2007b.WHO guidelines on Good Manufacturing Practices (GMP) for herbal medicines. Geneva: World Health Organization. p72.
- World Health Organization. 2011. Food safety and food borne illnesses; fact sheet. p237.
- World Health Organization. 2019. WHO global report on traditional and complementary Medicine 2019. Geneva. World Health Organization.
- Yesuf, A., Wondimeneh, Y., Gebrecherkos, T. and Moges, F. 2016. Occurrence of potential bacterial pathogens and their antimicrobial susceptibility patterns isolated from herbal medicinal products sold in different markets of Gondar town, Northwest Ethiopia. Int J Bacteriol. 2016:1959418.



How to cite this article:

Omoruyi, Z, Okundaye M. I. and Egunjobi, T. O. (2023). Evaluation of the microbial quality and safety of regulated and unregulated liquid herbal preparations in Benin City, Nigeria. Int. J. Adv. Res. Biol. Sci. 10(2): 59-71. DOI: http://dx.doi.org/10.22192/ijarbs.2023.10.02.008