International Journal of Advanced Research in Biological Sciences ISSN: 2348-8069 www.ijarbs.com

(A Peer Reviewed, Referred, Indexed and Open Access Journal) DOI: 10.22192/ijarbs Coden: IJARQG (USA) Volume 12, Issue 1-2025

Research Article



DOI: http://dx.doi.org/10.22192/ijarbs.2025.12.01.005

Antifeeding and repellent effect of plant extract Salvia officinalis against Tribolium castaneum (Herbst) (Coleoptera:Tenebrionidae)

Medona, L., Dharani, P., Krishnappa K., K. Elumalai*

Department of Zoology, Government Arts College (Autonomous), (Affiliated to the University of Madras), Nandanam, Chennai- 600035, Tamil Nadu, India *Corresponding Author E-mail: *professorelumalai@gmail.com*

Abstract

In the present investigation, we have tested the antifeedant, insecticidal and repellent activities of different solvent extracts of ethyl acetate and ethanol in *Salvia officinalis* Linn (Lamiaceae) against the selected red flour beetle, *Tribolium castaneum* Herbs. (Coleoptera: Tenebrionidae).The research findings showed that the solvent extracts had many activities. However, the ethanol extract had the highest mortality compared to ethyl acetate of *Salvia officinalis*. Thus, the isolation and identification of the phytochemical group found in the selected plant's ethanol extract are now underway. This present study showed that chemical pesticides would gradually replace phytochemicals.

Keywords: Salvia officinalis, Tribolium castaneum, Antifeedant activity, Repellent activity, ethanol, and ethyl acetate.

1.0 Introduction

In 2050, the global population is expected to exceed 9.1 billion, and food production is expected to increase by 70% to nourish this growing population (United Nations. World Population Prospects 2017). The seriousness of "food insecurity" highlights the enormous challenge of providing everyone with protected, nutrient-rich, and adequate food (WHO 2019). Unfortunately, increased agricultural production alone will not be enough to achieve the "zero hunger target" by 2030 or meet rising food demand.

Pre-harvest and post-harvest issues and insect infestation represent a significant restriction in optimal food production, resulting in massive grain losses. Before reaching the consumer, food grains undergo various processes, such as harvesting time, cleaning, drying, storage, processing, and transportation. It has been

determined that food losses in the post-harvest chain start at harvest and continue until food marketing at the consumer's end (Hodges et al., 2011; Mesterházy et al., 2020; Grolleaud, 2002). Technical limitations, including poor infrastructure, faulty packing, and inadequate inventory management systems, can also result in grain losses. Poring grains and food products is a significant problem worldwide, and insect pests cause controlled damage (Haq et al., 2005). Approximately 9% of stored grain yield is reduced in a favourable environment due to insect pests and mites (Fields, 2006).

Tribolium castaneum (Herbst) is the most serious pest of stored products known as the red flour beetle. It is included in the family Tenebrionidae, i.e. darkling beetles and red flour beetles, and is a common inhabitant of milled cereal products, stored flour, and fungus-infested grain (Park 1962; Beeman 2003; Campbell and Hagstrum 2002; McKay et al., 2019). T. castaneum causes damage to flour mills and other locations where dried foods and cereal products are stored or processed. T. castaneum adult females lay eggs on flour, complete their life cycle, and deplete the nutritional quality of grains over time. The flour becomes contaminated with a pungent smell in severe infestations and loses nutritional and market value (Payne 1925; A.S 1976). T. castaneum contaminates food products through moulting and excretion in addition to direct feeding, making the product commercially undesirable. The grain may be rejected depending on the level of infestation. Infested grains can have a profound impact on human health as well as a negative impact on the environment.

The red flour beetle is quite possibly the main put-away grain bug that has been accounted for to harm an assortment of products such as grains (Atta *et al.*, 2020), flour (Naseri *et al.*, 2017), peas (Pretheep-Kumar *et al.*, 2007), beans (Abdullahi *et al.*, 2018), nuts (Pires *et al.*, 2017), dried fruits (Sarwar, 2015) and spices (Tripathi *et al.*, 2009). Preventive and therapeutic control measures are used to control stored grain pests. Highly toxic synthetic pesticides have been used for many years among these. However, these chemicals have serious consequences for public health and the environment and insecticide resistance (Madkour *et al.*, 2012).

studies Several assessed that leaves S.officinalis are the source of insecticidal compounds to identify that rich content of bioactive compound molecules such as flavonoids, alkaloids, polyphenols, and tannins (Iqbal et al., 2006; Sharma et al., 1998), shows the source for new biopesticides (Rajashekar et al., 2014). These botanical insecticides have several advantages over synthetic ones, including environmental safety, less hazard, costeffectiveness, and easy availability (Gupta and Pathak, 2009; Tembo et al., 2018).

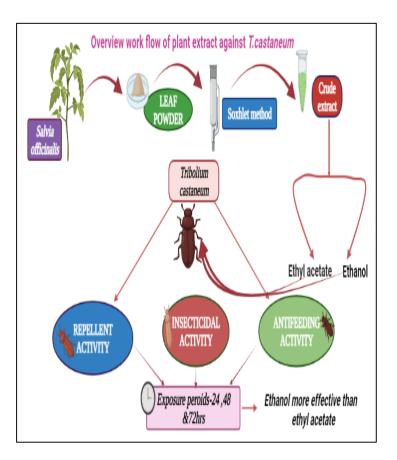


Fig1 Overview concept of work

The goal of this current study is to evaluate the effectiveness of *Salvia officinalis* ethanol and ethyl acetate extracts against the red flour beetle, *T. castaneum*, as well as to detect the antifeeding, insecticidal and repellent properties of various solvent extracts.

2.0 Materials and Methods

The research was conducted at the Department of Zoology in Government Arts College Nandanam. Insects were collected from the grains in the nearby local market. The insects are tested in a glass beaker under laboratory conditions. The beakers were closed with muslin cloth and tied with a rubber band to avoid the discharge of insects.

2.1 preparation of plant extract:

The fresh leaves of *S. officinalis* were collected from Kerala. Then, the leaves were dried shade at room temperature $(28\pm1^{\circ}C)$ for about 15 days. The dried leaves were powdered in the electric blender and stored in polyethene bags. The powder was packed in filter paper, and then the extract was extracted in the Soxhlet apparatus at 1:5 (plant powder: solvent) in ethyl acetate and ethanol solvent. After eight hours of extraction, the extract was kept in rotatory evaporation for 48 hrs. Then, extracts were stored at 4 °C in the refrigerator.

2.2 Rearing of *T.castaneum*:

T. castaneum was reared on sooji in our laboratory. The culture was maintained in the dark at $28 \pm 2^{\circ}$ C and $60 \pm 5\%$ RH. All adults used in the experiments were 1-10 days old. In the experiments, the day of death of the adult beetles was determined as the day the antennae and legs did not move upon gently disturbed using forceps.

2.3 Anti-feeding characteristic of crude extract against *T.castaneum*:

The probability of the antifeeding activity of the plant extracts was determined by Talukder & Howse (1993). Sooji flour was used as treated

feed. The sooji flour was sprayed with two different solvent extracts, air-dried for 30 minutes, and then weighed 200 gmof sooji in each box after 20 weevils were placed on them. The experiment was carried out by two different methods choice method and no-choice methods. In this choice method, feed with the prospect between treated (T) and control(C). In this nochoice method feed without the prospect of treated (TT) samples. Sooji was sprayed with three various periods: 20, 40, and 60 seconds. After 2 days again sooji flour was reweighed. The experiment was designed in a randomized complete pattern with five replications. Finally, the weight loss of sooji was estimated and the amount of feed consumed (FC) was also calculated.

 $FC = IW - [(FW \times IW)/FW]$

FC= feed consumed IW= initial weight of sooji ; FW= final weight of sooji

The amount of feed consumed in two different tests (TT and CT) was calculated:

Absolute coefficient of deterrence A= (TT - CT/TT + CT) \times 100

Relative coefficient of deterrence R= (C - T/C + T) \times 100

The total coefficient of deterrence T = A + R

2.4 Repellent characteristic of crude extract against *T.castaneum*:

Five plastic boxes serve as the basis of the bioassay system (8cm 5cm). These five boxes are divided into four for treatment and one for control. Plastic tubes with a 12 cm tube connecting their lumens were used to connect the five boxes. After ten pairs of unsexed adults were introduced in the insect chamber box. The different concentrations of 200ppm, 400ppm, 600ppm, and 800ppm. Neem Azal was for the control treatment. Five replications were repeated.

After 24, 48, and 72 hrs, control and treated boxes were calculated by using Lwande's method

$$EPI = \frac{Nt - Nc}{Nt + Nc} X \, 100$$

Nc- Number of insects in control sample Nt- Number of insects in treated sample

2.5 Insecticidal characteristic of crude extract against *T.castaneum*:

The insecticidal activity was performed on the newly emerged adults against *T.castaneum*. The Whatman no.1 filter paper was treated with

different concentrations (100, 200, 300, and 400 ppm) of different plant extracts and was allowed to dry for 10 minutes. Then, the filter paper was attached to the cap of the lid internally and in each plastic jar, twenty adult weevils were introduced in each box. Control was treated with Neem Azal. The insecticidal activity was calculated by using Abbott's formula.

$$POD = \frac{Ts - Cs}{Cs} X100$$

POD: Percentage of damage Ts: Number of insects in treated sample. Cs: Number of insects in control.

3.0 Results

Table 3.1 Repellent activity of ethyl acetate extract of S. officinalis tested against T. castaneum

Concentration	Exposure periods (in Hrs)		
	24 Hrs	48Hrs	72Hrs
200 ppm	13.5 ± 0.42^{a}	21.2 ± 1.47^{a}	27.6 ± 0.46^{a}
400 ppm	33.8±0.79 ^b	$40.2{\pm}0.55^{a}$	52.3±0.84 ^b
600 ppm	60.7 ± 1.12^{b}	64.3±0.71 ^a	68.7±1.33 ^c
800 ppm	$79.2 \pm 0.97^{\circ}$	82.8 ± 0.69^{b}	$94.7{\pm}0.88^{\circ}$
Neem azal	$100.0\pm0.00^{\text{ d}}$	$100.0\pm0.00^{\rm d}$	$100.0\pm0.00^{\rm d}$

Values expressed are mean mortality \pm standard deviations of five replications (n=20). Values with a different alphabet in the column show statistical significance at P<0.01% level.

The repellent activity of *S. officinalis* was tested against the stored pest of *T.castaneum* and the data obtained in the experiment are shown in table 3.1 and figure 3.1. It was observed that the highest repellent activity in the maximum concentration 800ppm in 24hrs (79.2 \pm 0.97) 48hrs (82.8 \pm 0.69) and 72hrs (94.7 \pm 0.88). In the same

induces24hrs way 600ppm concentration $((60.7\pm1.12)$ 48hrs (64.3 ± 0.71) and 72hrs (68.7±1.33). Likewise, 24hrs (33.8±0.79) 48hrs (40.2 ± 0.55) and 72hrs (52.8±0.84) and 48hrs (21.2±1.47) and (13.5 ± 0.42) 72hrs (27.6±0.46) mortality were observed from the concentrations of 400 and 200ppm respectively.

Table 3.2 Repellent activity	v of ethanol extract of S.or	fficinalis tested against	T.castaneum
	, .		

Concentration		Exposure periods (in Hrs)		
	24 Hrs	48Hrs	72Hrs	
200 ppm	17.7 ± 0.56^{a}	19.3±1.26 ^a	22.6 ± 0.38^{a}	
400 ppm	31.4 ± 0.99^{b}	$47.4{\pm}0.85^{a}$	55.3 ± 0.64^{b}	
600 ppm	66.1±1.31 ^b	61.3 ± 0.92^{a}	$67.9 \pm 1.43^{\circ}$	
800 ppm	84.5±1.87 ^c	89.9±1.04 ^b	96.2±1.86°	
Neem azal	$100.0\pm0.00^{\rm d}$	$100.0\pm0.00^{\rm d}$	$100.0\pm0.00^{\text{ d}}$	

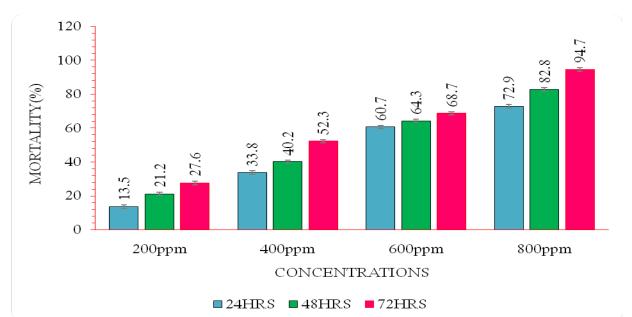
Values expressed are mean mortality \pm standard deviations of five replications (n=20). Values with a different alphabet in the column show statistical significance at P<0.01% level.

Int. J. Adv. Res. Biol. Sci. (2025). 12(1): 44-50

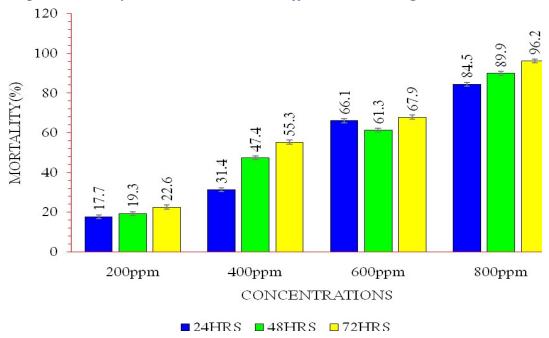
The repellent activity of *S. officinalis* was tested against the stored pest of *T. castaneum* and the data obtained in the experiment are shown in table 3.2 and Figure 3.2. It was observed that the highest repellent activity in the maximum concentration 800ppm in 24hrs (84.5 ± 1.87) 48hrs (89.9 ± 1.04) and 72hrs (96.2 ± 1.86).In the same

600ppm concentration induces 24hrs wav $((66.1 \pm 1.31))$ 48hrs (61.3 ± 0.92) & 72hrs 48hrs (67.9±1.43). Likewise (31.4 ± 0.99) (47.4±0.85) & 72hrs (55.3±0.64) and (17.7±0.56) 48hrs (19.3±1.26) & 72hrs (22.6±0.38) mortality were observed from the concentrations of 400 and 200ppm respectively.









Solvents	Exposure time	Coefficients of deterrence		Efficacy of	
Ethyl acetate	(min)	Absolute	Relative	Total	S. officinalis
	20	27.41	40.21	67.62c	+
	40	44.62	53.67	98.29bc	++
	60	57.01	84.46	141.47abc	+++
Ethanol	20	32.09	59.30	91.39c	+
	40	54.67	73.42	128.09bc	++
	60	78.41	97.87	176.28abc	+++

Table 3.3 Antifeeding deterrence coefficients of crude extract of *S.officinalis* tested against *T.castaneum*

The analysis of the antifeedant effect of *S. officinalis* shows that ethanol extract was more significant than ethyl acetate. Similarly, results of antifeedant effect on *S. oryzae* and *T. castaneum* were observed by Talukder & Howse (1993, 1994) with four different extracts of *Aphanamixis*

polystachya, acetone extract being the most significant with a total deterrence coefficient of 159.5. Valladares *et al.* (2003) also indicated an antifeedant effect of ethanol extract of senescent leaves of *Melia azedarach* on *S. oryzae*, with an antifeedant index of 100%.

Table 3.4 Insecticidal activity of ethyl acetate extract of S. officinalis tested against T. castaneum

Conc	Exposure periods (in Hrs)		
	24 Hrs	48 Hrs	72 Hrs
200 ppm	15.7 ± 0.32^{a}	$18.2{\pm}0.67^{a}$	$20.5{\pm}0.58^{a}$
400 ppm	29.8±0.86 ^b	31.9±0.95 ^a	42.7±0.91 ^b
600 ppm	58.5 ± 0.62^{b}	64.2±1.41 ^a	78.1±1.42 ^c
800 ppm	89.2±1.97°	92.3±1.89 ^b	$97.0{\pm}0.88^{\circ}$
Neem azal	100.0±0.00 ^d		
Values expressed are mean mortality \pm standard deviations of five replications (n=20). Values with the different alphabet in the column show statistical significance at P<0.01% level			

The insecticidal activity of *S. officinalis* was tested against the stored pest of *T. castaneum*, and the data obtained in the experiment are shown in Table 3.4 and Figure 3.3. It was observed that the highest repellent activity in the maximum concentration 800ppm in 24hrs (89.2 ± 1.97), 48hrs (92.9 ± 1.89), and 72hrs (97.0 ± 0.88).Similarly

600ppm concentration induces24hrs $(58.5.5\pm0.62)$ 48hrs (64.2±1.89)&72hrs (78.1±1.42). 24hrs In the same manner (29.8 ± 0.86) 48hrs (31.9 ± 0.95) & 72hrs (42.7±0.91) and (15.7±0.32) 48hrs (18.2±0.67) & 72hrs (20.5±0.58) mortality were observed from the concentrations of 400 and 200 ppm respectively

Table 3.5 Insecticidal activity of ethanol extract of S.officinalis tested against T.castaneum

Conc	Exposure periods (in Hrs)			
	24 Hrs	48Hrs	72Hrs	
200 ppm	16.5 ± 0.26^{a}	19.9±0.22 ^a	22.6 ± 0.63^{a}	
400 ppm	31.4±0.89 ^b	43.6±0.55 ^a	$62.4{\pm}0.97^{b}$	
600 ppm	67.5±1.06 ^b	$74.7{\pm}0.86^{a}$	88.7±129 ^c	
800 ppm	89.8±1.57 ^c	92.3±1.49 ^b	$98.5 \pm 1.88^{\circ}$	
Neem azal	100.0±0.00 ^d			
Values expressed are mean mortality \pm standard deviations of five replications (n=20). Values				
with the different alphabet in the column show statistical significance at P<0.01% level				

The insecticidal activity of *S. officinalis* was tested against the stored pest of *T. castaneum*, and the data obtained in the experiment are shown in Table 3.5 and Figure 3.4. It was observed that the highest repellent activity in the maximum concentration 800ppm in 24hrs (89.8 \pm 1.57), 48hrs (92.3 \pm 1.04), and 72hrs (98.5 \pm 1.88). Similarly,

600ppm concentration induces24hrs (67.5 ± 1.06) 48hrs (74.7 ± 0.86) & 72hrs (88.7 ± 1.29). In the same manner (31.4 ± 0.89) 48hrs (43.6 ± 0.55) & 72hrs (62.4 ± 0.97) and (16.5 ± 0.26) 48hrs (19.9 ± 0.22) & 72hrs (22.6 ± 0.63) mortality were observed from the concentrations of400 and200ppm respectively



Fig 2. Represent the lab activity

4.0 Discussion

Protecting crops from insects is one of the most critical global issues. Insects were commonly controlled with artificial insecticides, whose toxicity puts the health of agricultural workers, animals, and food consumers at risk. Because of the detrimental impacts on human health and their negligible costs and environmental consequences, plant-based pesticides have grown in popularity. This article discusses the anti-insect properties of beneficial plant substances (such as essential oils, flavonoids, alkaloids, glycosides, esters, and fatty acids) and their importance as a substitute for chemicals used in insect removal in a variety of ways, such as growth retardants, chemosterilants, repellents, antifeedants, toxicants, and attractants. Therefore, using botanical insecticides as an integrated insect management program can prominently reduce the use of artificial insecticides (Wafaa et al., (2017).

Atta *et al.*, (2020) assessed that the greatest mortality was reached with the highest dose of crude extract of *Z. officinale* at 10 days exposure interval. The findings of this study supported the use of *Z. officinale* because of its poisonous and repellent properties, which ultimately improve the grain quality of stored grains, in lowering the population of *T. castaneum*.

Many plant species produce a wide range of chemical compounds that can be toxic, deterring, or repellant to insect pests. Some of these substances are also toxic to plants, so they are kept in particular organs like seeds and flowers. These substances are specially designed to combat pest insects that feed on plants. (Shojaaddini *et al.*, 2008). Monoterpenes could be one of the most effective and safest alternatives to artificial insecticides (Popovic *et al.*, 2013). Monoterpenes can enter the insect's respiratory system and rapidly interact with physiological functions. Similar to the effects of pyrethroid insecticides, these substances can also directly stimulate as neurotoxic substances, affecting acetylcholinesterase activity and causing hyperactivity, convulsions, and tremors.

According to Pugazhvendan et al. (2009), stored food grains suffer severe damage from insect infestation. The losses range from 5 to 30% of total agricultural output worldwide. T. castaneum has been discovered in various commodities, including cereals, flour, peas, nuts, dried figs, and spices. It infests groundnut kernels and pods as well. Due to its high reproductive potential, this insect causes significant loss in storage and can breed all year in warm areas. Synthetic pesticides are discouraged due to their adverse effects on human health and the environment. The maximum repellent activity was noticed in T.purprea powder, with EPI values of -0.11 and -0.56 at 1 and 6 hours against T. castaneum, respectively.

Likewise, Nujira Tatun *et al.* (2014) studied the extracts of *R. communis* and *C. papaya*, which were controlled in nutrition, improved larval mortality, and extended the intervals of the larval and pupal periods. The ratios of pupated and emerging adults and the amount of F1 offspring were decreased by feeding them. Also, Mostafa *et al.*, 2012 showed that four plant extracts revealed a strong to moderate toxicity at different concentrations on red flour beetle. *Cucumis sativus* extracts have the highest mortality among leaf extracts, whereas *Psidium guajava* extract showed the lowest mortality rate (50%).

Extracts of *Fumeria indicia*, *Viola odorata*, and *Linium statism* at various percentage doses were applied for their significant effects on insect mortality and population growth rate. After the application of revealed that *V. odorata* extract significantly displayed higher insect mortality, lower population growth rate levels, and the least reduction in grain weight (Waqar Islam and others, 2016).

Similarly, the repellent activity of clove, coriander, neem, and mint extracts against red flour beetle destroys storage grains and other

The findings of this investigation crops. demonstrated that these plant extracts have insect-repelling properties. successful It concluded that these plants are natural sources of repellent material and are a potential source of natural/biological insect repellents (Abid Ramsha et al., 2019)-the efficacy of several medicinal plant extracts to kill T. castaneum, which infests stored goods. According to the findings, A. sativum (garlic) and Z. Officinale (ginger) were more active, causing a 15 times increased adult death rate and a 4 to 5 times reduction in grain weight losses. Surprisingly, either the lowest or highest doses showed improved control. The findings of this study suggest the use of botanicals for pest management of stored merchandise (Faheem Ahmed et al., 2019).

Likewise, plant oil has a high potential to be used as a bio-pesticide in managing *the T. castaneum* pest (Jeyasankar and Cheeaiyan 2016). Additionally, Agarwal and Agasha (2020) analyzed the microbiome of a crucial red flour beetle, a widespread pest of stored cereals. The beetle was a complete flour lifecycle, including habitat, food, and microbes. Finally, under various tested conditions, the microbiome was identified as not essential for survival and growth. So, it was concluded that *Tribolium castaneum* closely mimicked the host species' natural niche.

Previously, researchers Shamjana and Grace 2022 studied T. castaneum resistance mechanisms and used bioassays and genetic and biochemical techniques. The use of whole-genome and gene expression analysis sequencing channels provided a more extensive collection of gene resources for future research into T. castaneum resistance mechanisms. whichinclude 1) target site insensitivity, 2) metabolic resistance, and 3) Less cuticular penetration. These various resistance mechanisms have been identified to act in layers to deteriorate the insecticides faster before they release their toxic effect, as shown in Fig 3.

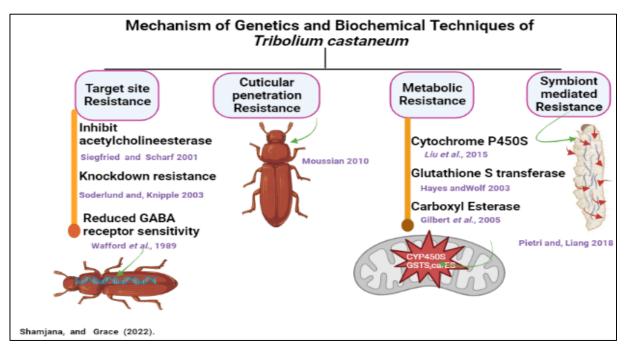


Figure-3-Mechanism of crude extracts against *T.castaneum*

Because of their persistent nature, these chemicals pose a significant environmental risk due to their increased risk of neurotoxic, carcinogenic, teratogenic, and genotoxicity effects in non-target animals, chronic residual toxicity, ability to cause hormonal imbalance, and hematotoxicity. (Khatter 2011). However, there is currently one alternative for protecting stored grains from insect damage. Plants are a good source of pest control compounds. The plant kingdom can be a rich source of chemicals with the potential for development as effective pest control agents.

Hence, it concluded that the effect of solvent extract of *S. officinalis* was tested on the stored beetles-*T. castaneum* and it was observed that the toxicity of solvent leaf extract was influenced by the solvent as well as the aromatic fragrance of the plant. The highest toxic effect was observed in the ethanol of *S. officinalis*. The highest mortality percentage was found to be directly proportional to the level of doses of plant extract. The powder of these botanicals may have also disrupted the respiratory activities of *T. castaneum*, leading to asphyxiation and death.

Conclusion

T. castaneum is one of the most prevalent and frequent pests in red flour and wheat flour in

various agricultural products. Farmers who store their belongings in containers will suffer a significant loss. For the study, two different solvents were used to extract the plant S. officinalis: ethanol and ethyl acetate. *S. officinalis* extracts from various solvents were tested at various concentrations, and their potency was observed and calculated. The best efficiency was found in the ethanol extracts from this seed, which have a high polarity index.

The current analysis indicates that plant-based substances are easily degradable and do not affect humans, animals, or the environment. So, *S. officinalis* could be used as the best and most potent natural agent to control the stored red flour pest of *Tribolium castaneum*.

Acknowledgments

The authors express sincere gratitude to The Principal, Government College Arts (Autonomous), Nandanam, Chennai and the Head of the Department, Department of Zoology Government (Autonomous), Arts College Nandanam. Chennai 600035. for their magnanimous concern.

References

- 1. Abid Ramsha, Khan Aisha Saleem and Butt Saba. (2019). Repellent activity of certain plant extracts (clove, coriander, neem and mint) against red flour beetle. *American Scientific Research Journal for Engineering, Technology, and Sciences (ASRJETS).* 55(1):83-91.
- 2. Abbott, W.S. (1925). A method of computing the effectiveness of insecticides. *Journal of Economic Entomology* 18: 265-267.
- 3. Agarwal A and Agashe D (2020). The red flour beetle *Tribolium castaneum*. A model for microbiome interactions. *Plos One* 15(10):1-26.
- 4. AS, A. (1976). Dhaliwal GS. Insect pests of stored grain and other Products. *Agricultural pests of India and South-East Asia. Kalyani Publisher, New Delhi, India*, 389-415.
- Atta, B., Rizwan, M., Sabir, A.M., Gogi, M.D. and Ali, K., (2020). The damage potential of *Tribolium castaneum* (Herbst) (Coleoptera: Tenebrionidae) on wheat grains stored in hermetic and non-hermetic storage bags. *International Journal of Tropical Insect Science*. 40: 27-37.
- Beeman, R. W. (2003). Distribution of the Medea factor M4 in *Tribolium castaneum* (Herbst) populations in the United States. *Journal of Stored Products Research*, 39(1), 45-51.
- Campbell, J. F., and Hagstrum, D. W. (2002). Patch exploitation by *Tribolium castaneum*: movement patterns, distribution, and oviposition. *Journal of Stored Products Research*, 38(1), 55-68.
- Faheem Ahmad *et* al. (2019). Comparative insecticidal activity of different plant materials from six common plant species against *Tribolium castaneum*. (Herbst) (Coleoptera: Tenebrionidae). *Saudi Journal of Biological Sciences* 26: 1804–1808.
- 9. Fields, P.G (2006). Effect of *Pisum sativum* fractions on nine stored-grain beetles' mortality and progeny production. *Journal of Stored Products Research*. 42: 86-96.

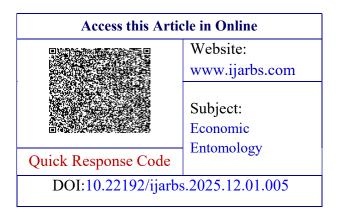
- 10. Gilbert, L. I., Iatrou, K., and Gill, S. S. (2005). *Comprehensive molecular insect science*. Elsevier.
- 11. Grolleaud, M. (2002). Post-harvest losses: discovering the whole story. Overview of the phenomenon of losses during the post-harvest system.
- 12. Gupta, M.P. and Pathak, R.K., (2009). Bioefficacy of neem products and insecticides against the incidence of whitefly, yellow mosaic virus and pod borer in black gram. *Natural Product Radiance*. 8(2): 133-136.
- Haq, T., Usmani, N.F. and Abbas, T (2005). Screening of plant leaves as grain protectants against *Tribolium castaneum* during storage. *Pakistan Journal of Botany* 37: 149-153.
- 14. Hodges, R. J., Buzby, J. C., and Bennett, B. (2011). Post-harvest losses and waste in developed and less developed countries: opportunities to improve resource use. *The Journal of Agricultural Science*, 149(S1), 37-45.
- 15. Jeyasankar, A. Chennaiyan V and T. Chinnamani. (2016). Evaluation of Five Essential Plant Oils as a Source of Repellent and Larvicidal Activities against Larvae of *Tribolium castaneum* (Herbst) (Coleoptera: Tenebrionidae). *Journal of Entomology* 13:98-103.
- 16. Khatter, N. A. (2011) Efficiency of Azardirachitin, a chitin inhibitor or growth, development and reproduction potential of *Tribolium confusum* after adult treatment. Journal of Entomology. 8(5): 440-449.
- 17. Madkour, M.H., Zaitoun, A.A. and Singer, F.A., (2012). Efficacy of three plant species extracts in controlling *Trogoderma granarium* Everts (Coleoptera: Dermestidae). *Journal of Food Agriculture and Environment*. 10: 1200-1203.
- McKay, T., Bowombe-Toko, M. P., Starkus, L. A., Arthur, F. H., and Campbell, J. F. (2019). Monitoring of *Tribolium castaneum* (Coleoptera: Tenebrionidae) in rice mills using pheromone-baited traps. *Journal of economic entomology*, *112*(3), 1454-1462.

- 19. Mesterházy, Á., Oláh, J., and Popp, J. (2020). Losses in the grain supply chain: Causes and solutions. *Sustainability*, *12*(6), 2342.
- 20. Mostafa M. *et al*, (2012). Insecticidal activity of plant extracts against *Tribolium castaneum* Herbst. J Adv Sci Res. 3(3): 80-84
- 21. Moussian, B. (2010). Recent advances in understanding mechanisms of insect cuticle differentiation. *Insect biochemistry and molecular biology*, 40(5), 363-375.
- 22. Naseri, B *et al.*, (2017). Influence of different food commodities on life history, feeding efficiency, and digestive enzymatic activity of *Tribolium castaneum* (Coleoptera: Tenebrionidae). *Journal of Economic Entomology*. 110(5): 2263-2268.
- 23. Nujira Tatun *et al*, (2014). Inhibitory effects of plant extracts on growth, development and α -amylase activity in the red flour beetle *Tribolium castaneum* (Coleoptera: Tenebrionidae). *Eur. J. Entomology.* 111(2): 181–188
- 24. Park, T. (1962). Beetles, Competition, and Populations: An intricate ecological phenomenon is brought into the laboratory and studied as an experimental model. *Science*, *138*(3548), 1369-1375.
- 25. Payne, N. M. (1925). Some Effects of Triboliumon on Flour. *Journal of Economic Entomology*, 18(5), 737-744.
- 26. Pietri J.E and Liang D.(2018). The link between insect symbionts and insecticide resistance: causal relationships and physiological trade-offs. *Annals of the Entomological society of America*. 111(3):92-97.
- 27. Popovic, A. Sucur, J. Orcic, D. and Strbac, P. (2013) Effect of essential oil formulations on the adult insect *Tribolium castaneum* (Herbst) (Coleoptera: Tenebrionidae). *Journal of Central European Agriculture*. 14(2): 181-193.
- Pretheep-Kumar, P., Mohan, S. and Ramaraju, K (2007). Long-term efficacy of proteinenriched pea flour against *Tribolium castaneum* (Coleoptera: Tenebrionidae) in wheat flour. *Journal of CentralEuropean Agriculture*. 7(4): 779-784.

- Pugazhvendan, S. R. Elumalai, K. Ross, P. R. and Soundararajan, M. (2009) Repellent activity of chosen plant species against *Tribolium castaneum*. World Journal of Zoology. 4(3): 188-190.
- 30. Rajendran S and Sriranjini V, (2008). Plant products as fumigants for stored-product insect control. *Journal of Stored Product Research*. 44: 126–135.
- Rehman *et al.* (2020). Compatibility of some botanicals and the entomopathogenic fungus, *Beauveria bassiana* (Bals.), against the red flour beetle, *Tribolium castaneum* (Herbst) (Coleoptera: Tenebrionidae). *Egyptian Journal of Biological Pest Control*.30:131.
- 32. Sarwar, M., (2015). Protecting dried fruits and vegetables against insect pest's invasions during drying and storage. *American Journal of Marketing Research*. 1(3): 142-149.
- 33. Shamjana, U., and Grace, T. (2022). Review of Insecticide Resistance and Its Underlying Mechanisms in *Tribolium castaneum*. Insecticides Impact and Benefits of Its Use for Humanity.
- 34. Shojaaddini, M., Moharramipour, S. and Sahaf, B.Z., (2008). Fumigant toxicity of essential oil from *Carum copticum* against Indian meal moth, *Plodia interpunctella*. Journal of Plant Protection Research. 48(4): 411-419
- 35. Siegfried, B. D., and Scharf, M. E. (2001). Mechanisms of organophosphate resistance in insects. In *Biochemical sites of insecticide action and resistance* (pp. 269-291). Springer, Berlin, Heidelberg.
- 36. Sies, H., and Ketterer, B. (Eds.). (1988). *Glutathione conjugation* (Vol. 748). London: Academic Press.
- 37. Soderlund, D. M., and Knipple, D. C. (2003). The molecular biology of knockdown resistance to pyrethroid insecticides. *Insect biochemistry and molecular biology*, *33*(6), 563-577.
- 38. Talukder F.A. & Howse P.E. (1993): Deterrent and insecticidal effects of extracts of pithraj, *Aphanamixis polystachya* (Meliaceae), against *Tribolium castaneum* in storage. Journal of Chemical Ecology, 19: 2463–2471.

- 39. Tembo, Y., Mkindi, A.G., Mkenda, P.A., Mpumi, N., Mwanauta, R., Stevenson, P.C., Ndakidemi, P.A. and Belmain, S.R., (2018). Pesticidal plant extracts improve yield and reduce insect pests on legume crops without harming beneficial arthropods. *Frontiers in Plant Science*. 9: 1425
- 40. Tripathi, A.K., Singh, A.K. and Upadhyay, S. (2009). Contact and fumigant toxicity of some common spices against the storage insects *Callosobruchusmaculatus* (Coleoptera: Bruchidae) and *Triboliumcastaneum* (Coleoptera: Tenebrionidae). *International Journal of Tropical Insect Science*. 29(3): 151-157.
- 41. Valladares, G., Defago, M. T., Palacios, S. M. and Carpinella, M. C. (1997). Laboratory evaluation of Melia azedarach (Meliaceae) extracts against the elm leaf beetle (Coleoptera: Chrysomelidae). *Journal of Economic Entomology*, 90: 747-750.

- 42. Wafaa M. *et al*, (2017). Botanical insecticide as simple extractives for pest control. *Cogent Biology*, 3:1.
- Wafford, K. A., Lummis, S. C. R., and Sattelle, D. B. (1989). Block of an insect central nervous system GABA receptor by cyclodiene and cyclohexane insecticides. *Proceedings of the Royal Society of London. B. Biological Sciences*, 237(1286), 53-61.
- 44. Waqar Islam (2016).Inhibitory Effects of Medicinal Plant Extracts Against *Tribolium castaneum* (Herbst.) (Coleoptera: Tenebrionidae). *Mayfeb Journal of Agricultural Science*3: 15-20.
- 45. World Health Organization. (2019). The state of food security and nutrition in the world 2019: safeguarding against economic slowdowns and downturns (Vol. 2019). Food and Agriculture Org...



How to cite this article:

Medona, L., Dharani, P., Krishnappa K., K. Elumalai. (2025). Antifeeding and repellent effect of plant extract *Salvia officinalis* against *Tribolium castaneum* (Herbst) (Coleoptera:Tenebrionidae). Int. J. Adv. Res. Biol. Sci. 12(1): 44-55. DOI: http://dx.doi.org/10.22192/ijarbs.2025.12.01.005