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Research Article

Effect of phosphate solubilizing bacteria and compost mixture on growth parameters in Paddy (*Oryza sativa* L.)

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Abstract

Phosphorus is one of the major essential macronutrients limiting plant growth owing to its low bioavailability in soils. The biofertilizer effect of phosphate solubilizing bacteria on growth parameters of *Oryza sativa* was studied in a field experiment. Maximum plant height was recorded during the harvest and highest plant height (121.98 cm) was recorded in the treatment T_8 (75% NPK + CM+ BS + BM + EA). Maximum dry matter production was recorded during the harvest and more dry matter production (8.12 g plant⁻¹) was recorded in the treatment T_8 . Maximum Leaf area index was recorded (6.42 cm) was recorded in the treatment T_8 .

Keywords: Leaf area index, Dry matter production, Plant height and Paddy.

Introduction

Phosphorus (P) is as an essential mineral nutrient for plant growth and development is the world's second highest chemical input in agriculture. Soluble P is often the limiting mineral nutrient for biomass production in agricultural ecosystems as well (Hameeda et al., 2006). Plants utilize fewer amounts of phosphate fertilizers that are applied and the rest is rapidly converted into insoluble complexes in the soil. So, this phenomenon encourages farmers to frequent application of phosphate fertilizers. Modern agriculture is severely modifying and polluting the natural environment, due to the widespread application of chemical fertilizers, herbicides and pesticides. Therefore, thinking about valid alternative for chemical fertilizers is too necessary. Phosphate solubilizing bacteria (PSB) are used as biofertilizer since 1950's (Kudashev, 1956; Krasilinikov, 1957; Saranraj et al., 2013; Sivasakthi et al., 2013). These microorganisms secrete different types of organic acids e.g., carboxylic acid (Deubel and Merbach,

2005;) thus lowering the pH in the rhizosphere (He and Zhu, 1988) and consequently dissociate the bound forms of phosphate like $Ca_3(PO_4)_2$ in calcareous soils. Use of these microorganisms as environment friendly biofertilizer helps to reduce the much expensive phosphatic fertilizers. Phosphorus biofertilizers could help increase the availability of accumulated phosphate (by solubilization), efficiency of biological nitrogen fixation and increase the availability of Fe, Zn etc., through production of plant growth promoting substances (Kucey et al., 1989; Kanchana et al., 2013). Trials with PSB indicated yield increases in rice (Tiwari et al., 1989; Usharani et al., 2013), maize (Pal, 1999) and other cereals (Ozturk et al., 2003; Afzal et al., 2005; Sivasakthi et al., 2014; Usharani et al., 2014; Kanchana et al., 2014).

Materials and Methods Treatment schedule

 $T_1 - 100\%$ NPK

T₂ – 75% NPK + CM + Bacillus subtilis (BS)

 $T_{3} - 75\% NPK + CM + Bacillus megaterium(BM)$ $T_{4} - 75\% NPK + CM + Enterobacter asburiae$ EA) $T_{5} - 75\% NPK + CM + BS + BM$ $T_{6} - 75\% NPK + CM + BM + EA$ $T_{7} - 75\% NPK + CM + EA + BS$ $T_{8} - 75\% CM + BS + BM + EA$ $T_{9} - Control$

Determination of growth parameters

Effect of plant height

The height of the plants in each treatment was measured at 30^{th} day after transplantation (DAT). The mean value of the plants from 3 replications was recorded.

Dry matter production

Plant samples were taken at three stages of crop growth viz., maximum tillering, flowering and harvest. The plant samples were dried in hot air oven at 80°C for 48 to 72 hours. The oven dry weight was recorded and Dry matter production was calculated.

Leaf area index

Leaf area index at flowering was calculated without removing the leaves by using the formula given by Yoshida *et al.* (1976).

LA	=	KxWxL Where,
LA	=	leaf area in cm
Κ	=	a constant factor (0.75)
W	=	maximum width of the 3 rd leaf from the top (in
cm)		
L	=	length of the 3 leaf from the top (in cm)

The leaf area of the middle tiller of the sample hill was calculated and was multiplied by the number of tillers in each hill. The: total leaf area arrived by multiplying the leaf area hill⁻¹ with number of hill m⁻¹ The LAI was computed using the formula given below.

LAI =
$$\frac{\text{Total leaf area per unit area } (\text{cm}^2)}{\text{Unit land area } (\text{cm}^2)}$$

Results and Discussion

The field experiment was conducted to study the effect sugar mill waste (pressmud) by phosphate solubilizing bacteria (*Bacillus subtilis, Bacillus megaterium* and *Enterobacter asburiae*) growth parameters of paddy (*Oryza sativa* L.) var BPT- 5804 grown rhizosphere soil. Phosphate solubilizing bacteria triple inoculants recorded maximum plant height. The observations recorded on plant height at 30 DAS, 60 DAS and at harvest are presented in Table – 1. Among the various treatments tested, the highest plant height was recorded in treatment T₈ (75% NPK + CM +BS+BM+EA) (121 .98 cm) and T₁ – 100% NPK (121.52 cm) were on par with each other. The minimum plant height was recorded in T₉ (Control) (89.30 cm).

The effect of sugar mill waste, Compost mixture (Pressmud + *Bacillus subtilis* + *Bacillus megaterium* + *Enterobacter asburiae*) chlorophyll content of paddy var BPT - 5804 (*Oryza sativa* L.) was investigated and the results were furnished in Table – 2. Among the nine treatments tested, the dry matter production was maximum in treatment T₈ (75% NPK + CM + BS + BM + EA) (8.12 t ha⁻¹) and T₁ – 100% NPK (7.93 t ha⁻¹) was on par with each other. The minimum plant height was recorded in T₉ (Control) (4.00 t ha⁻¹).

The effect of sugar mill waste, Compost mixture (Pressmud + *Bacillus subtilis* + *Bacillus megaterium* + *Enterobacter asburiae*) Leaf area index at flowering of paddy var BPT - 5804 (*Oryza sativa* L.) was investigated and the results were furnished in Table – 3. Among the nine treatments tested, the Leaf area index at flowering was maximum in treatment T₈ (75% NPK + CM+BS+BM+EA) (6.42 cm) and T₁ – 100% NPK (6.02cm) was on par with each other. The minimum plant height was recorded in T₉ (Control) (3.98 cm).

Enhancement of seedling growth due to seed treatment with phosphate solubilizing bacteria like B. subtilis and B. megaterium may be due to release of plant growth promoting substances. Plant growth promoting an microbes are important contributor to biofertilization of agricultural crops. Production of growth regulators by phosphate solubilizing bacteria has been studied by Ponmurugan and Gopi (2006).There is increasing evidence that phosphobacteria improve plant growth due to biosynthesis of plant growth substances rather than their action in releasing available phosphorus. Chaykovskava et al. (2001) reported that treatment

Tuestmente	Plant height (cm)		
1 reatments	30 DAS	60 DAS	Harvest
T ₁ – 100% NPK	55.37	82.02	121.52
$T_2 - 75\%$ NPK + CM + Bacillus subtilis (BS)	50.12	72.60	110.20
T_3 - 75% NPK + CM + Bacillus megaterium (BM)	50.83	73.99	112.86
T_4 -75% NPK + CM + Enterobacter asburiae (EA)	49.93	70.02	109.07
$T_5 - 75\%$ NPK + CM+BS+BM	54.81	78.12	119.83
$T_6 - 75\%$ NPK + CM+BM+EA	53.78	76.91	119.71
$T_7 - 75\%$ NPK + CM+EA+BS	52.91	75.80	118.36
$T_8 - 75\% + CM + BS + BM + EA$	56.13	82.70	121.98
T ₉ - Control	27.80	50.55	89.30
SE _D	0.41	0.35	0.24
CD (P=0.05)	0.83	0.71	0.48

Int. J. Adv. Res. Biol.Sci. 1(9): (2014): 263–267 Table – 1: Plant height of paddy (*Oryza sativa L*.) cv –BPT 5804

Table – 2: Dry matter production of Paddy (*Oryza sativa L.*) var BPT 5804

Treatments	Dry matter production (t ha ⁻¹)
T ₁ - 100% NPK	7.93
$T_2 - 75\%$ NPK + CM + Bacillus subtilis (BS)	6.43
T ₃ - 75% NPK + CM + Bacillus megaterium (BM)	6.93
T ₄ -75% NPK + CM + Enterobacter asburiae (EA)	6.00
T ₅ - 75% NPK + CM+BS+BM	7.68
$T_6 - 75\%$ NPK + CM+BM+EA	7.45
$T_7 - 75\%$ NPK + CM+EA+BS	7.12
$T_8 - 75\%$ CM+BS+BM+EA	8.12
T ₉ - Control	4.00
SE _D	0.51
CD (P=0.05)	1.03

Table – 3: Leaf area index at flowering (Oryza sativa L.) cv –BPT 5804

Treatments	Leaf area index at flowering (cm)
T ₁ - 100% NPK	6.02
$T_2 - 75\%$ NPK + CM + Bacillus subtilis (BS)	5.00
T ₃ - 75% NPK + CM + Bacillus megaterium (BM)	5.12
T_4 -75% NPK + CM + Enterobacter asburiae (EA)	4.89
$T_5 - 75\%$ NPK + CM+BS+BM	5.98
T ₆ - 75% NPK + CM+BM+EA	5.77
T ₇ - 75% NPK + CM+EA+BS	5.32
$T_8 - 75\%$ CM+BS+BM+EA	6.42
T ₉ - Control	3.98
SE _D	0.4
CD (P=0.05)	0.8

with phosphate solubilizing bacteria resulted in increased yield of pea and barley. Several results suggest that phosphate-solubilizing bacteria have the ability to solubilize phosphate thereby increasing its availability to plants. Goenadi *et al.* (2000) suggested that the inoculation of PSB could enhance the maximum growth as well as result increased productivity. Among the PSB, will be more effective in increasing growth with inoculation of PSB will give better results as compared to control (Kanchana *et al.*, 2014). Hence, use of these phosphate-solubilizing bacteria, as biofertilizer should be promoted (Sivasakthivelan and Saranraj, 2013; Kanchana *et al.*, 2013; Usharani *et al.*, 2014).

References

- Afzal, A., M. Ashraf, S.A. Asad, M. Farooq 2005. Effect of phosphate solubilizing microorganisms on phosphorus uptake, yield and yield traits of wheat (*Triticum aestivum* L.) in rainfed area. Int. J Agric. Biol. 7:207-9.
- Chaykovskaya, L.A., V.P. Patyka and T.M. Melnychuk, 2001. Phosphorus mobilizing microorganisms and their influence on the productivity of plants. In: Plant Nutrition-Food Security and Sustainability of Agroecosystem. Horst WJ (Ed) pp: 668-669.
- Deubel A. W. Merbach . 2005. Influence of microorganisms on phosphorus bioavailability in soils. In: Buscot, F. and A. Varma (eds.), Microorganisms in Soils: Roles in Genesis and Functions. pp. 177-191.
- Goenadi, D.H., Y. Siswanto and Y. Sugiarto, 2000. Soil science society of America Journal, 64: 927-932.
- Hameeda, B., G. Harmi, O.P. Rupela, S.P. Wani and G. Reddy, 2006. Growth promotion of maize by phosphate solubilising bacteria isolated from composts and macrofauna. Microbiol, 59: 23-147.
- He Z.L, J. Zhu .1988). Microbial utilization and transformation of phosphate adsorbed by variable charged minerals. Soil Biol. Biochem. 30:917-923.
- Kanchana, D., M. Jayanthi, D. Kanchana, P. Saranraj and D. Sujitha. 2013. Evaluation of Plant growth promoting substance production by *Azospirillum* sp. isolated from rhizosphere of Chilli (*Capsicum annuum* L.). *International Journal of Microbiological Research*, 4(3): 300 304.
- Kanchana, D., M. Jayanthi, G. Usharani, P. Saranraj and D. Sujitha. 2013. Prevalence of *Azotobacter* sp.

in Chilli (*Capsicum annuum* L.) rhizosphere soil of Cuddalore district, Tamil Nadu, India. *International Journal of Microbiological Research*, 4(3): 296 - 299.

- Kanchana, D., M. Jayanthi, G. Usharani, P. Saranraj and D. Sujitha. 2014. Interaction effect of combined inoculation of PGPR on growth and yield parameters of Chili var K1 (*Capsicum* annuum L.). International Journal of Microbiological Research, 5(3): 144 - 151.
- Krasilinikov, N.A .1957. On the role of soil microorganism in plant nutrition. Microbiologiya 26:659-672.
- Kucey RMN, Janzen HH, Leggett ME (1989). Microbially mediated increases in plant-available phosphorus. Ad Agron. 42:199-228.
- Kudashev I.S .1956. The effect of phosphobacterin on the yield and protein content in grains of Autumm wheat, maize and soybean. Doki Akad Skh Nauk. 8:20-23.
- Ozturk, A. O. Caglar, F. Sahin .2003. Yield response of wheat and barley to inoculation of plant growth promoting rhizobacteria at various levels of nitrogen fertilization. J. Plant Nutr. Soil Sci. 166:1-5.
- Pal, S. 1999). Interaction of an acid tolerant strain of phosphate solubilizing bacteria with a few acid tolerant crops. Plant Soil 213:221-230.
- Ponmurugan, P. and C. Gopi, 2006. African J Biotechnol, 5(4): 348-350
- Saranraj, P., P. Sivasakthivelan and S. Siva Sakthi. 2013. Prevalence and production of plant growth promoting substance by *Pseudomonas fluorescens* isolated from paddy rhizosphere soil of Cuddalore district, Tamil Nadu, India. *African Journal of Basic and Applied Sciences*, 5 (2): 95 - 101.
- Sivasakthi, S., D. Kanchana, G. Usharani and P. Saranraj. 2013. Production of plant growth promoting substance by *Pseudomonas fluorescens* and *Bacillus subtilis* isolated from paddy rhizosphere soil of Cuddalore district, Tamil Nadu, India. *International Journal of Microbiological Research*, 4(3): 227 233.
- Sivasakthi, S., G. Usharani and P. Saranraj. 2014. Biocontrol potentiality of Plant growth promoting rhizobacteria (PGPR) – *P. fluorescens* and *B. subtilis*: A Review. *African Journal of Agricultural Sciences*, 9(16): 1265 – 1277.
- Sivasakthivelan, P and P. Saranraj. 2013. *Azospirillum* and its formulations: A Review. *International*

Journal of Microbiological Research, 4(3): 275 - 287.

- Tiwari V.N., L.K. Lehri, A.N. Pathak. 1989. Effect of inoculating crops with phospho-microbes. Exp. Agric. 25:47-50.
- Usharani, G., D. Kanchana, M. Jayanthi, P. Saranraj and D. Sujitha. 2013. Evaluation of certain resistance inducing chemicals against Sheath blight incidence in Paddy (*Oryza sativa* L.). *International Journal of Microbiological Research*, 4(3): 333 -335.
- Usharani, G., D. Kanchana, M. Jayanthi, P. Saranraj and D. Sujitha. 2014. Effect of Salicylic acid and *Pseudomonas fluorescence* on the growth and yield of Paddy IR-50. *International Journal of Microbiological Research*, 5(1): 54 - 60.
- Usharani, G., D. Sujitha, S. Sivasakthi and P. Saranraj. 2014. Effect of Arbuscular Mycorrhizal (AM) Fungi (*Glomus fasciculatum* L.) for the improvement of growth and yield of Maize (*Zea mays* L.). *Central European Journal of Experimental Biology*, 3(2): 19 – 25.
- Usharani, G., P. Saranraj, D. Sujitha and S. Sivasakthi. 2014. Effect of *Azotobacter* sp. for the improvement of growth and yield of Pearl Millet (Cumbu) (*Pennisetum glaucum* L.). *International Journal of Advanced Multidisciplinary Research*, 1(4): 102 – 106.
- Usharani, M. Jayanthi, D. Kanchana and P. Saranraj. 2014. Effect of *Bradyrhizobium* isolates for the maximization of growth and yield of Black gram (*Vigna mungo* L.). *International Journal of Advanced Multidisciplinary Research*, 1(4): 33 – 39.